

Bioactive secondary metabolites from fruiting bodies of Higher Fungi

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- **Introduction**
- **Secondary metabolites from fruiting bodies of *Hygrophorus***
- **Secondary metabolites from fruiting bodies of *Cortinarius***
- **Bioactivity of fungal secondary metabolites**

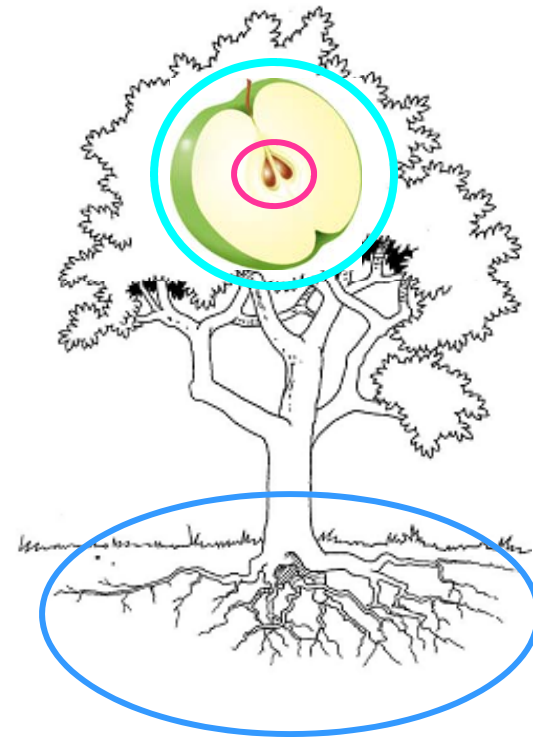
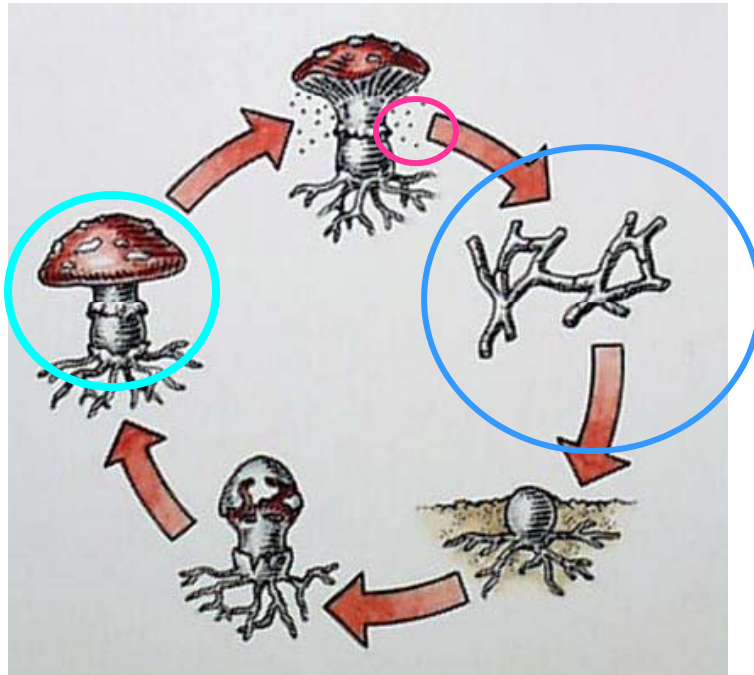


Fungal fruiting bodies as a source of natural products

Why dealing with fungi ?

- Evolutionary very old organism
- First multicellular organisms which are successfully growing on soil
- Life cycle: different environment, different enemies





fungi: most of the life cycle (mycelium) under the grassroot

reproductive organs (fruiting bodies): ~ 1 month over the grassroot

e.g.: *Armillaria bulbosa*: 1 500 years old, over 10 000 kg, 15 ha (Nature 356: 428-431)

Armillaria ostoyae: 2 400 years old, 900 ha



Fungal fruiting bodies as a source of natural products

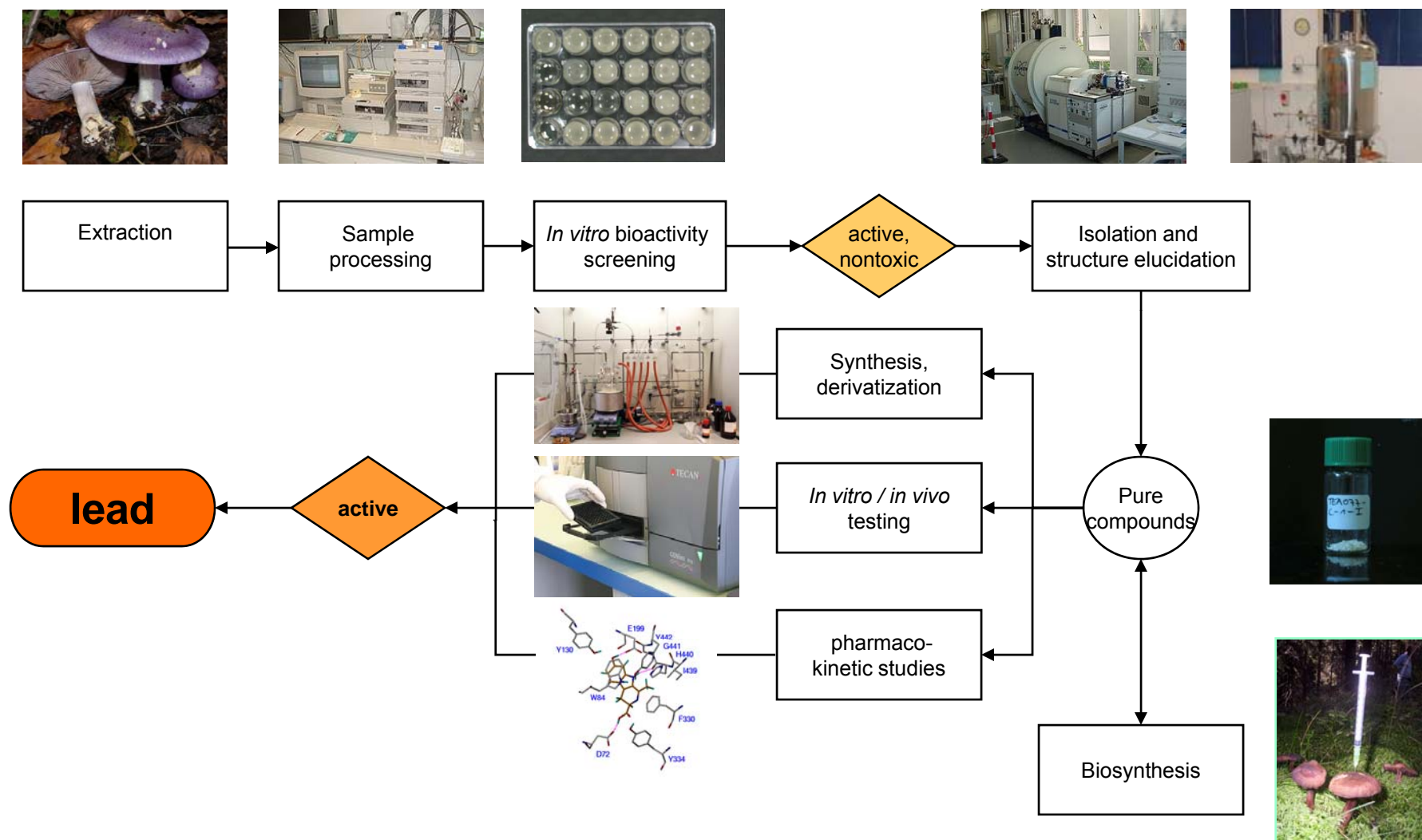
Why dealing with fungi ?

- Evolutionary very old organism
- First multicellular organisms which are successfully growing on soil
- Life cycle: different environment, different enemies
- High specialisation (ectomycorrhiza, endomycorrhiza, endophytes)

➔ **wide spectrum of new compounds expected**



Bioassay guided isolation of secondary metabolites



Hygrophorones A – G from *Hygrophorus* spp. Sect. *Olivaceoumbrini*



H. personii



H. olivaceoalbus



H. latitabundus



H. pustulatus



Genus *Hygrophorus*

- engl.: wax caps
 - Tricholomataceae (Basidiomycetes)
 - ~ 60 species
 - symbionts (mycorrhiza) with trees
 - fruiting bodies strong slimy surface
 - various colour reactions of the stipes
 - **field observation:**
rarely attacked by parasites (pathogenic fungi, yeasts,...)
- **assumption: bioactive secondary metabolites as preform defense**



Hygrophorus in Europe

Sect. **Hygrophorus** – Subsect. Chrysodonti

H. chrysodon

Gelbzahn-
Schneckling



Sect. **Hygrophorus** – Subsect. Pallidini

H. penarius

Trockener
Schneckling



Sect. **Hygrophorus** – Subsect. Hygrophorus

H. eburneus

Elfenbein-
Schneckling



H. hedrychii

Birkenschneckling



H. carpini

Hainbuchen-
Schneckling



H. cossus

Verfärbender
Schneckling



H. gliocyclus

Schleimbringter
Schneckling



H. chryaspis



Sect. **Pudorini** – Subsect. Erubescentes

H. erubescens

Rasiger
Purpurschneckling



H. russula

Purpur-
schneckling



11 *H. capreolarius*

Weinroter
Schneckling



Sect. **Pudorini** – Subsect. Pudorini

H. poetarum

Isabellrötlicher
Schneckling



H. nemoreus

Wald-Schneckling



H. pudorinus

Orange-Schneckling



Sect. **Discoidei**

H. discoideus

Braunscheibiger
Schneckling



H. unicolor

Orangefalber
Schneckling



H. lucorum

Lärchen-
Schneckling



H. hypothejus

Frostschneckling



Sect. **Olivaceoumrini** – Subsect. – Olivaceoumrini

H. olivaceoalbus

Natterstieler
Schneckling



H. persoonii

Olivgestiefler
Schneckling



H. latitabundus

Großer Kiefern-
Schneckling



Sect. **Olivaceoumrini** – Subsect. – Tephroleuci

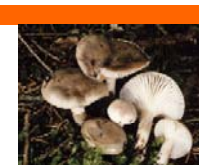
H. pustulatus

Schwarzpunktierter
Schneckling



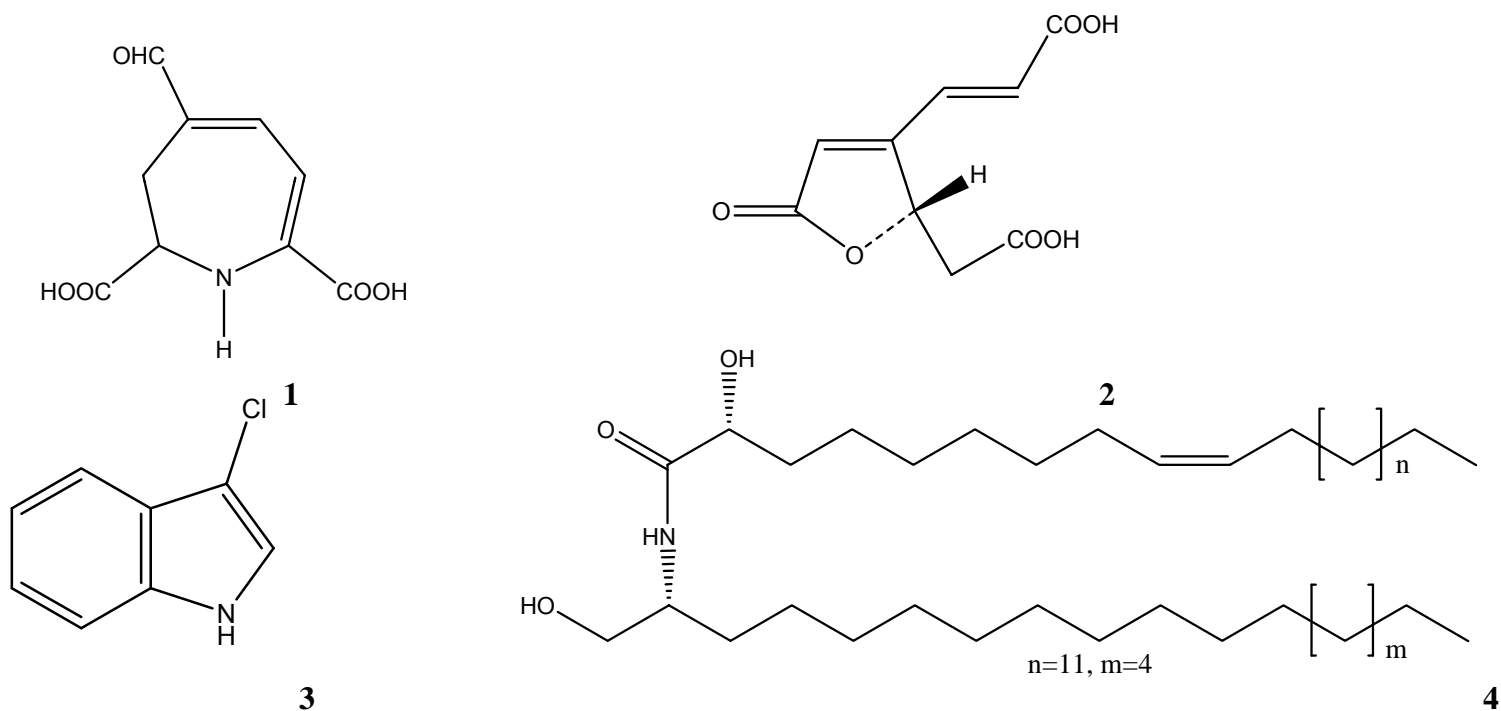
H. agathosmus

Mehliger
Schneckling



Previous works on metabolites from *Hygrophorus*

- Fugmann et al. (1985): muscaflavin (1) and hygrophoric acid (2) in *H. aureus*, *H. hypothejus*, *H. lucorum* and *H. speciosus*
- Wood et al. (2003): 3-Chloroindole (3) in *H. paupertinus*
- Qu et al. (2004): Hygrophamide (4) in *H. eburneus*



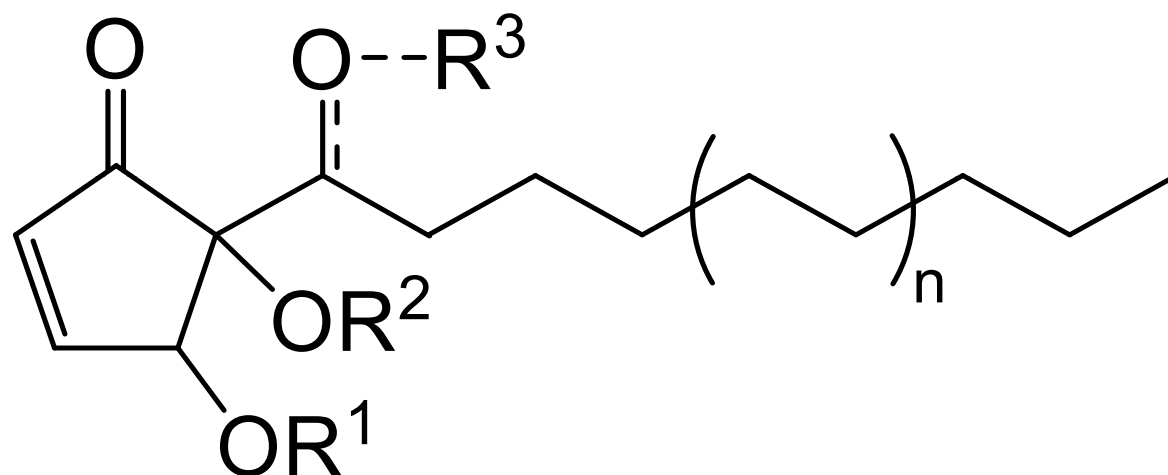
Fugmann, B. (1985) – Neue niedermolekulare Naturstoffe aus Höheren Pilzen. Dissertation, Universität Bonn

Wood, W., Smith, J., Wayman, K., Largent, D. (2003) – Indole and 3-chloroindole: the source of the disagreeable odor of *Hygrophorus paupertinus*. *Mycologia* 95(5), 807 – 808

Qu, Y., Zhang, H., Liu, J. (2004) – Isolation and Structure of a new ceramide from the Basidiomycete *Hygrophorus eburneus*. *Z. Naturforsch.* 59b, 241 - 244



Hygrophorones A – G from *Hygrophorus* spp.

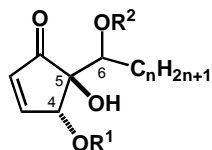


Lübken, T., Schmidt, J., Porzel, A., Arnold, N., & Wessjohann, L. (2004) - Hygrophorones A–G: fungicidal cyclopentenones from *Hygrophorus* species (Basidiomycetes). *Phytochemistry*, 65, 1061-1071.

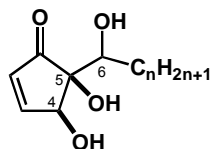
Lübken, T., Arnold, N. & Wessjohann, L. Hygrophorone und deren Derivate. – PCT/EP/05001957



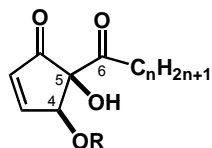
Hygrophorones A – G from *Hygrophorus* spp.



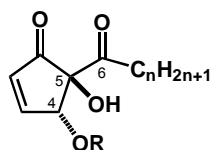
- 10 4,6-Di-O-acetyl-Hygrophoron A¹² R¹ = Ac R² = Ac n = 12 *H. persoonii*
 11 4-O-Acetyl-Hygrophoron A¹² R¹ = Ac R² = H n = 12 *H. persoonii*
 12 6-O-Acetyl-Hygrophoron A¹² R¹ = H R² = Ac n = 12 *H. persoonii*
 13 4,5-Di-O-acetyl-Hygrophoron A¹⁴ R¹ = Ac R² = Ac n = 14 *H. persoonii*
 14 4-O-Acetyl-Hygrophoron A¹⁴ R¹ = Ac R² = H n = 14 *H. persoonii*
 15 6-O-Acetyl-Hygrophoron A¹⁴ R¹ = H R² = Ac n = 14 *H. persoonii*



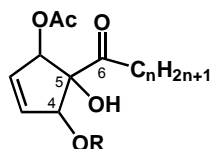
- 16 Hygrophoron B¹⁴ n = 14 *H. olivaceoalbus*
 17 Hygrophoron B¹⁶ n = 16 *H. olivaceoalbus*



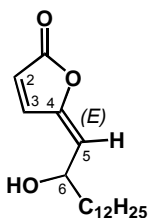
- 18 4-O-Acetyl-Hygrophoron C¹² R = Ac n = 12 *H. pustulatus*
 19 Hygrophoron C¹² R = H n = 12 *H. pustulatus*



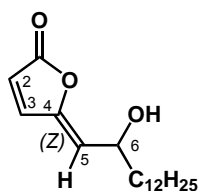
- 20 4-O-Acetyl-Hygrophoron D¹² R = Ac n = 12 *H. latitabundus*
 21 Hygrophoron D¹² R = H n = 12 *H. latitabundus*
 22 4-O-Acetyl-Hygrophoron D¹⁴ R = Ac n = 14 *H. latitabundus*



- 23 1,4-Di-O-acetyl-Hygrophoron E¹² R = Ac n = 12 *H. latitabundus*
 24 1,4-Di-O-acetyl-Hygrophoron E¹⁰ R = Ac n = 10 *H. latitabundus*
 25 1,4-Di-O-acetyl-Hygrophoron E¹⁴ R = Ac n = 14 *H. latitabundus*
 26 1-O-Acetyl-Hygrophoron E¹² R = H n = 12 *H. latitabundus*
 27 1-O-Acetyl-Hygrophoron E¹⁰ R = H n = 10 *H. latitabundus*



- 28 Hygrophoron F¹²
H. persoonii



- 29 Hygrophoron G¹²
H. persoonii



H. persoonii



H. olivaceoalbus



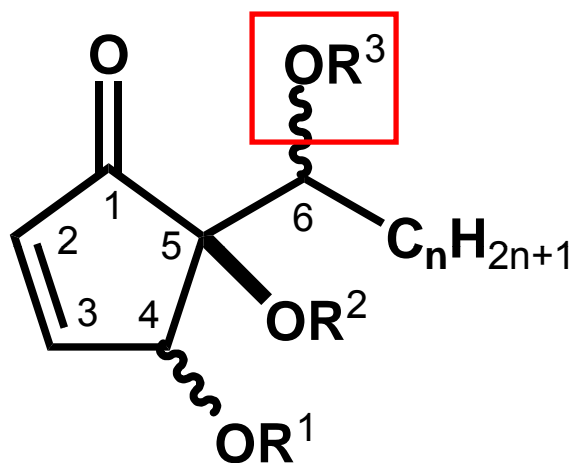
H. pustulatus



H. latitabundus

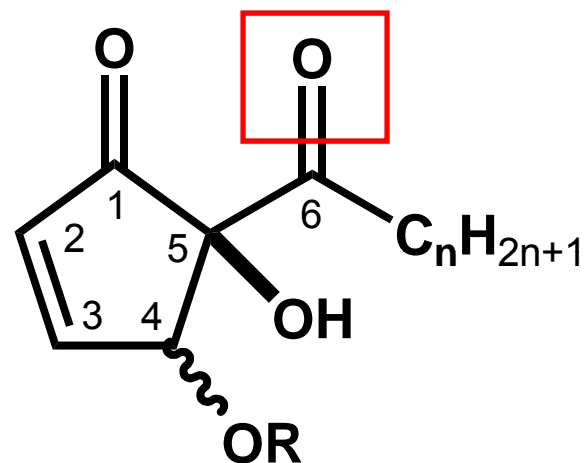


Structures of Hygrophorones



R¹, R², R³ = H, Ac

Type I



R = H, Ac

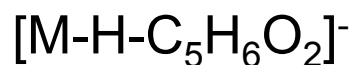
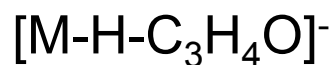
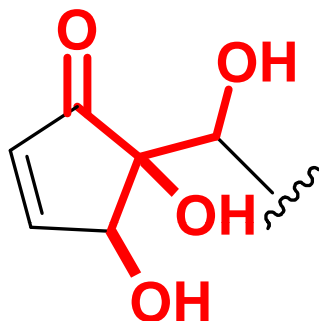
Type II

Lübken, T., Arnold, N., Wessjohann, L., Böttcher, C. & Schmidt, J. (2006) - Analysis of fungal cyclopentenone derivatives from *Hygrophorus* spp. by liquid chromatography/electrospray-tandem mass spectrometry. *J. Mass Spectrometry* 41 (3), 361 – 371.

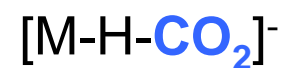
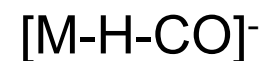
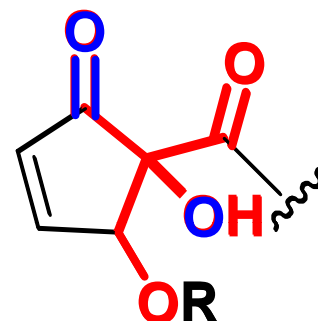


Relation between structural features and negative ion ESI-CID mass spectra

Hygrophorones Type I



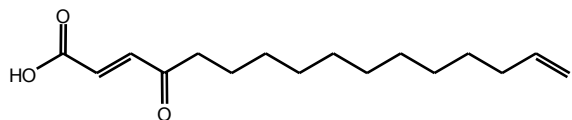
Hygrophorones Type II



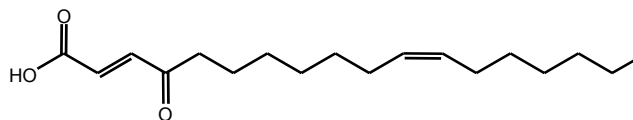
Lübken, T., Arnold, N., Wessjohann, L., Böttcher, C. & Schmidt, J. (2006) - Analysis of fungal cyclopentenone derivatives from *Hygrophorus* spp. by liquid chromatography/electrospray-tandem mass spectrometry. *J. Mass Spectrometry* 41 (3), 361 – 371.



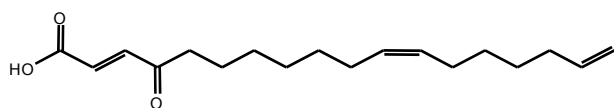
4-Oxo-2-alkenoic Fatty Acids



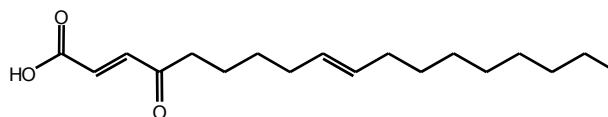
(E)-4-oxohexadeca-2,15-dienoic acid



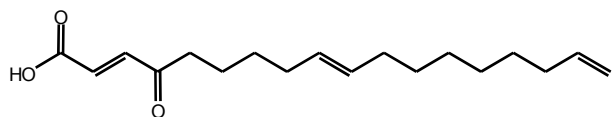
(2E,11Z)-4-oxooctadeca-2,11-dienoic acid



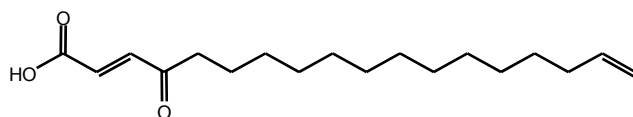
(2E,11Z)-4-oxooctadeca-2,11,17-trienoic acid



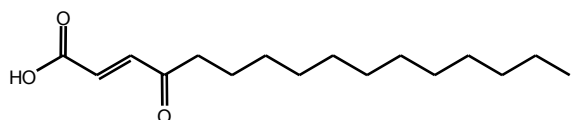
(2E,9E)-4-oxooctadeca-2,9-dienoic acid



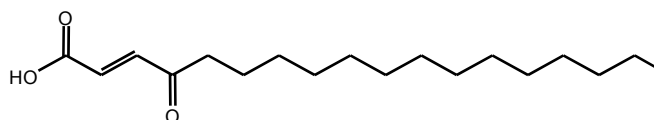
(2E,9E)-4-oxooctadeca-2,9,17-trienoic acid



(E)-4-oxooctadeca-2,17-dienoic acid



(E)-4-oxohexadeca-2-enoic acid



(E)-4-oxooctadeca-2-enoic acid



H. eburneus

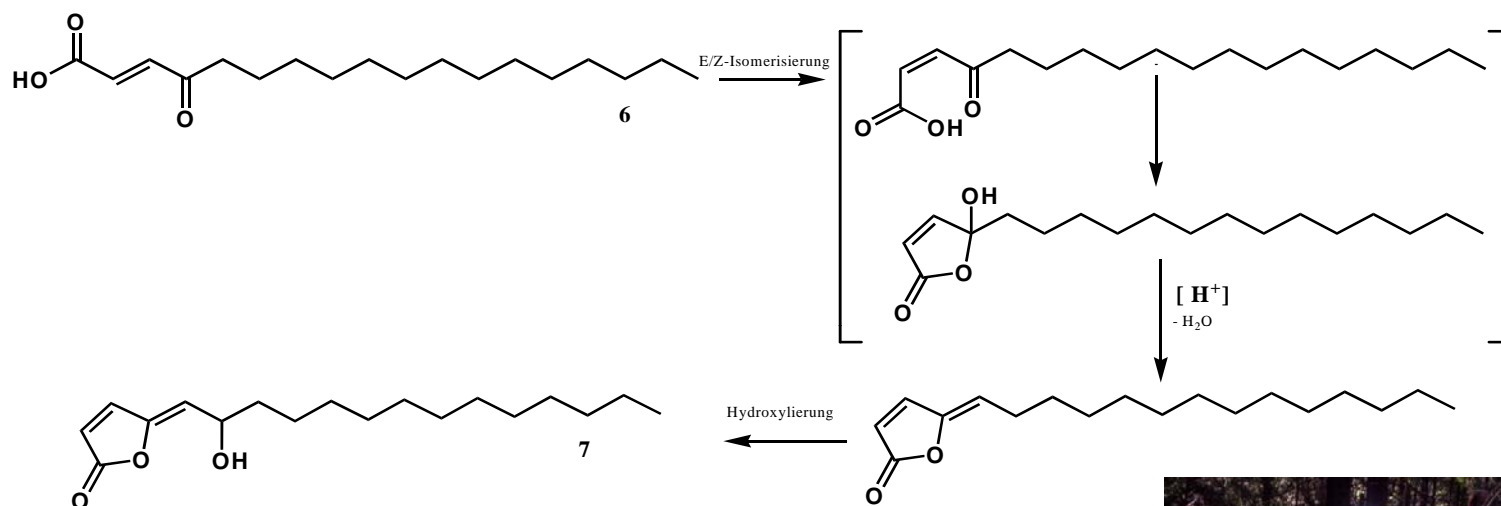
Teichert, A., Lübken, T., Schmidt, J., Porzel, A., Arnold, N. & Wessjohann, L. (2005) - Unusual bioactive 4-oxo-2-alkenoic fatty acids from *Hygrophorus eburneus*. Z. Naturforsch. 60b, 25 - 32

Teichert, A., Lübken, T., Kummer, M., Besl, H., Haslberger, H. & Arnold, N. (2005) - Bioaktive Sekundärmetaboliten aus der Gattung *Hygrophorus* (Basidiomycetes). Z. Mykol., 71/1, 53 - 62



Proposed biosynthesis

(6) potential biosynthetic precursor of Hygrophoron G¹² (7) ?



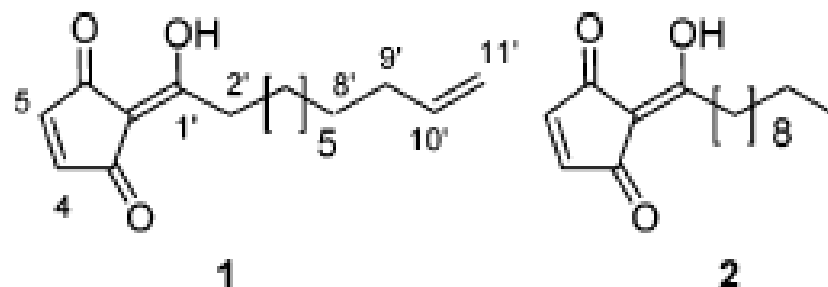
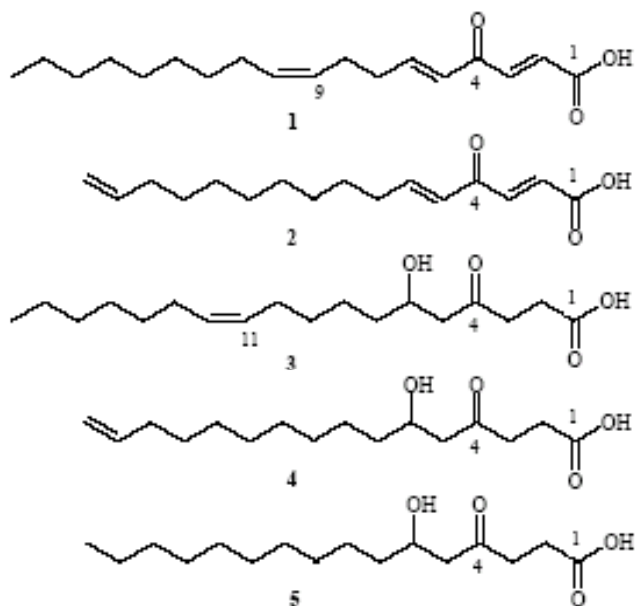
Feeding experiments with synthetic ¹³C labelled fatty acids
→ **no incorporation detectable** (low concentration, weather conditions,...)



Group Vidari (2006, 2007)

4-Oxo Fatty Acids from *Hygrophorus discoxanthus*

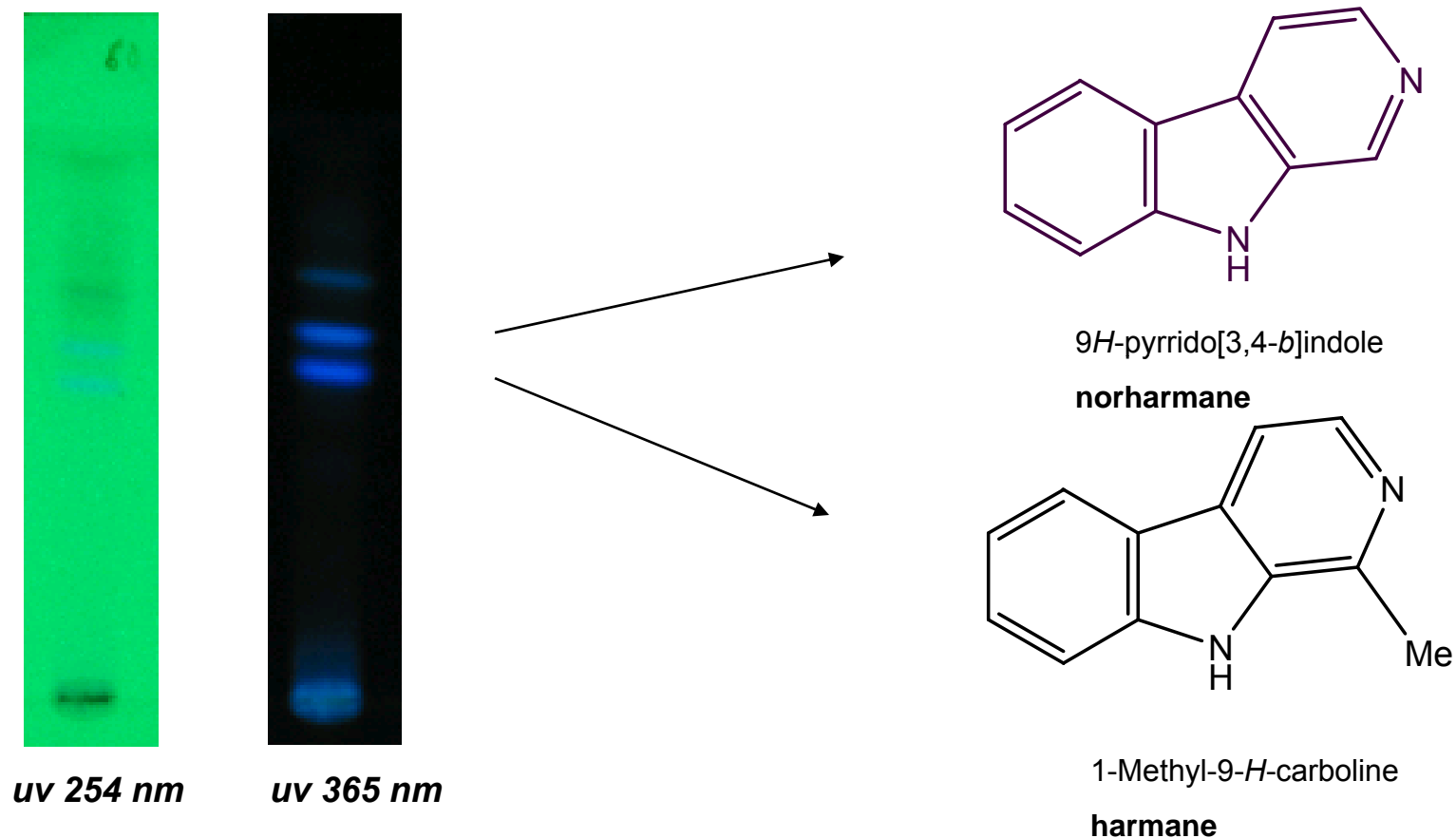
Acylcyclopentenediones from of *Hygrophorus chrysodon*



Gianluca Gilardoni, Marco Clericuzio, Solveig Tosi, Giuseppe Zanoni, and Giovanni Vidari (2007) - Antifungal Acylcyclopentenediones from Fruiting Bodies of *Hygrophorus chrysodon* *J. Nat. Prod.*, 70, 137-139
Gilardoni, G.; Clericuzio, M.; Marchetti, A.; Vita Finzi, P.; Zanoni, G.; Vidari, G. *Nat. Prod. Commun.* 2006, New oxidized 4-Oxo Fatty Acids from *Hygrophorus discoxanthus*. *Nat. Prod. Commun.*, 1, 1079-1084.



Occurrence of β -carboline alkaloids harmane and norharmane in *Hygrophorus* spp.

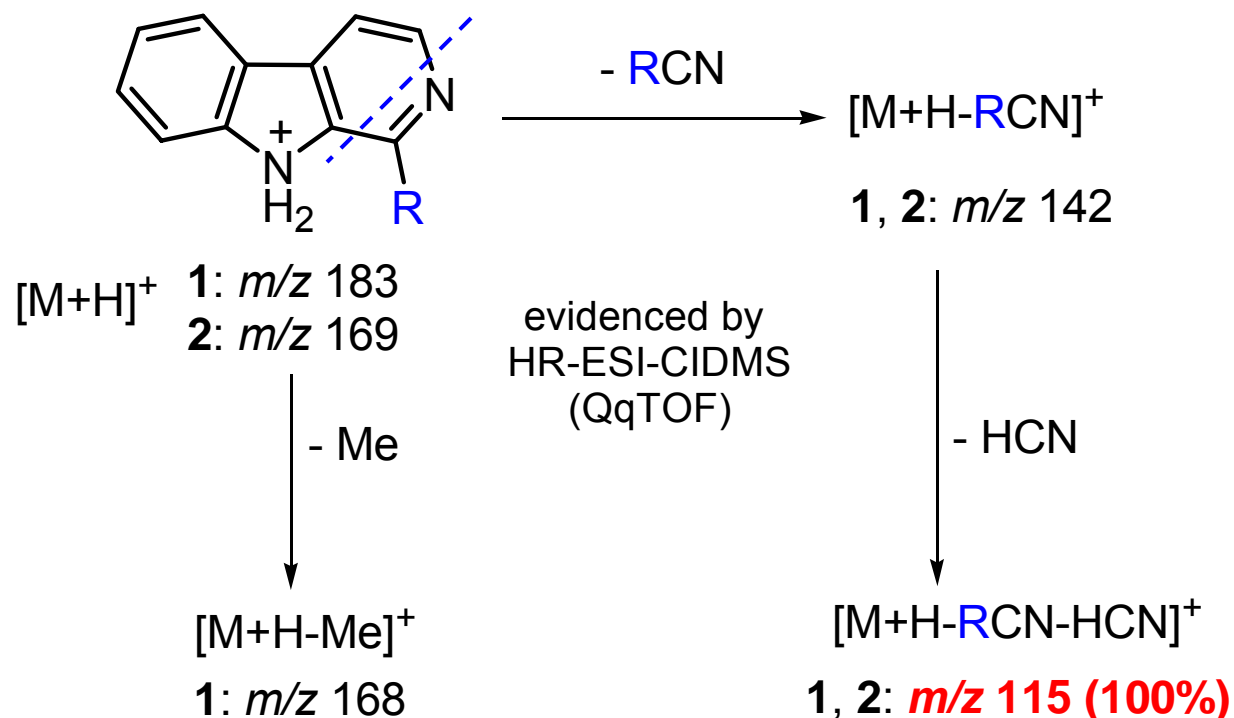


Axel Teichert, Å., Schmidt, J., Kuhnt, C., Huth, M., Porzel, A., Wessjohann, L., Arnold, N. (2008) - Determination of β -carboline alkaloids in fruiting bodies of *Hygrophorus* spp. by liquid chromatography/electrospray ionisation tandem mass spectrometry. *Phytochem. Analysis.*, in press.



Main Fragmentation of Harmane and Norharmane

- 1: Harmane (R=Me)
2: Norharmane (R=H)

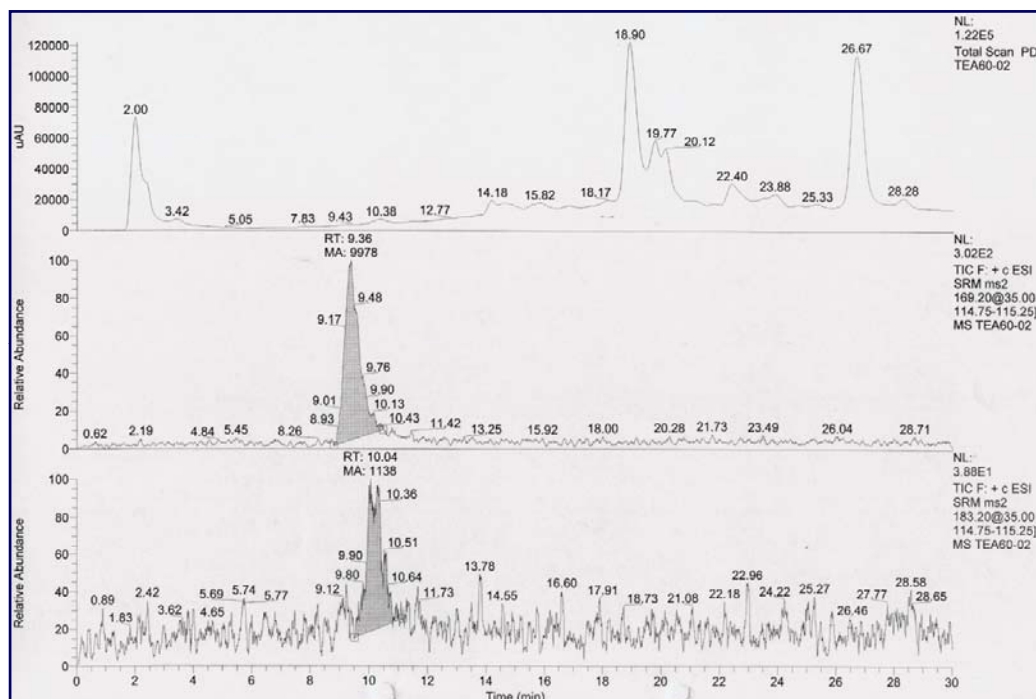


Axel Teichert, Å., Schmidt, J., Kuhnt, C., Huth, M., Porzel, A., Wessjohann, L., Arnold, N. (2008) - Determination of β -carboline alkaloids in fruiting bodies of *Hygrophorus* spp. by liquid chromatography/electrospray ionisation tandem mass spectrometry. *Phytochem. Analysis.*, in press.



Evidence for the co-occurrence of harmane and norharmane in 28 investigated *Hygrophorus* species by LC/ESI-SRM

Species	Norharmane pg μL^{-1}	Harmane pg μL^{-1}
<i>H. agathosmus</i>	1,04	0,30
<i>H. carpinii</i>	5,81	2,71
<i>H. chrysoaspis</i>	3,06	4,85
<i>H. chrysodon</i>	15,84	0,94
<i>H. cossus</i>	16,98	1,25
<i>H. dichrous</i>	0,07	0,07
<i>H. discoideus</i>	31,04	2,31
<i>H. discoxanthus</i>	0,82	1,62
<i>H. eburneus</i>	95,62	8,62
<i>H. erubescens</i>	1,85	1,42
<i>H. fuscoalbus</i>	0,60	0,23
<i>H. glyocyclus</i>	0,91	0,86
<i>H. hedrychii</i>	3,82	0,65
<i>H. hyacinthinus</i>	3,44	5,47
<i>H. hypothejus</i>	13,16	1,68
<i>H. latitabundus</i>	8,96	1,83
<i>H. lucorum</i>	2,61	1,92
<i>H. marzuolus</i>	2,56	2,43
<i>H. nemoreus</i>	0,89	1,22
<i>H. olivaceosalbus</i>	3,81	0,42
<i>H. penarius</i>	8,50	5,63
<i>H. persoonii</i>	46,46	11,10
<i>H. poetarum</i>	4,13	0,83
<i>H. pudorinus</i>	0,54	0,85
<i>H. pustulatus</i>	2,39	3,91
<i>H. russula</i>	0,48	0,64
<i>H. speciosus</i>	0,78	0,53
<i>H. unicolor</i>	0,98	0,79



→ harmane and norharmane - a chemotaxonomic marker

Axel Teichert, Å., Schmidt, J., Kuhnt, C., Huth, M., Porzel, A., Wessjohann, L., Arnold, N. (2008) - Determination of β -carboline alkaloids in fruiting bodies of *Hygrophorus* spp. by liquid chromatography/electrospray ionisation tandem mass spectrometry. *Phytochem. Analysis.*, in press.



***Hygrophorus hyacinthinus* Quel.**
(very rare species)



daylight



uv light (365 nm)



Isolation

MeOH crude extract



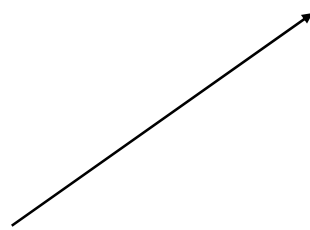
Diaion



Sephadex LH20



prep. HPLC, RP18



MeOH
365 nm



TLC 365 nm

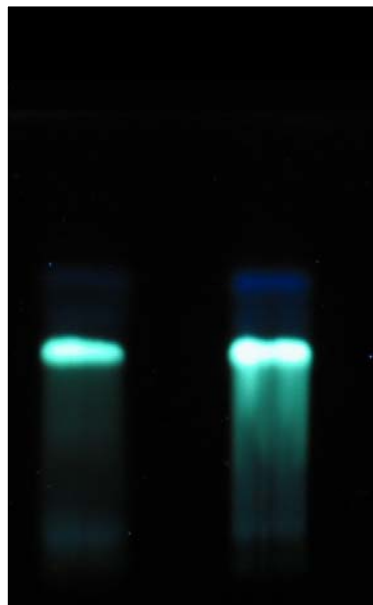
HR-MS: m/z 245.092 (calc. for $C_{13}H_{13}O_3N_2$, $[M+H]^+$, 245.092)

m/z 201.102 (calc. for $C_{12}H_{13}O_1N_2$, $[M+H]^+$, 201.102) \rightarrow $[M+H-CO_2]^+$



Similarities to *Cortinarius brunneus* – identical substances!

H. hyacinthinus



Cortinarius brunneus

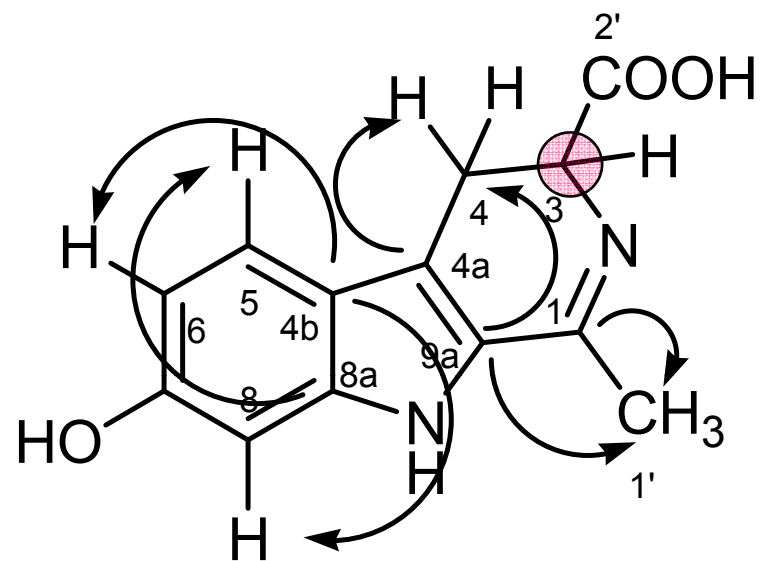


TLC: BuOH:HOAc:H₂O, 4:1:1

Teichert, A., Schmidt, J., Porzel, A., Arnold, N., & Wessjohann, L. - Brunneins A – D, β -carboline alkaloids from *Cortinarius brunneus* (Basidiomycetes). *J. Nat. Prod.*, 70(9), 1529-1531.



^{13}C NMR



Brunnein A

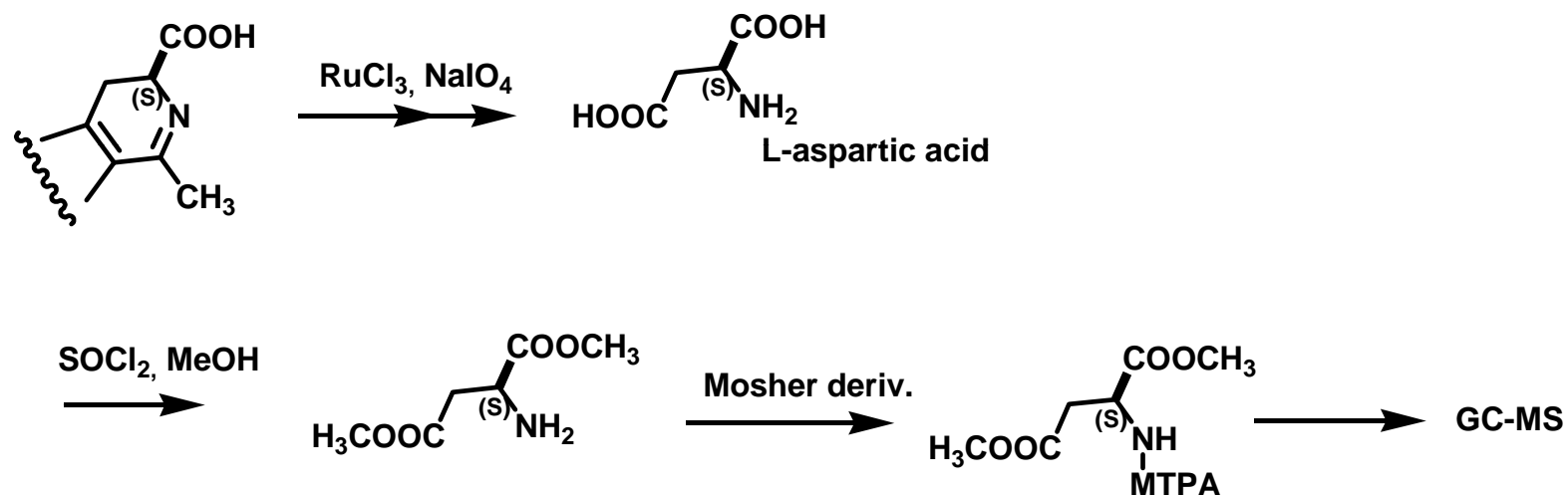
13 carbons: *i.a.* 6 quaternary C

1 chiral center \rightarrow H-3_{ax}



Brunnein A: configuration at C-3

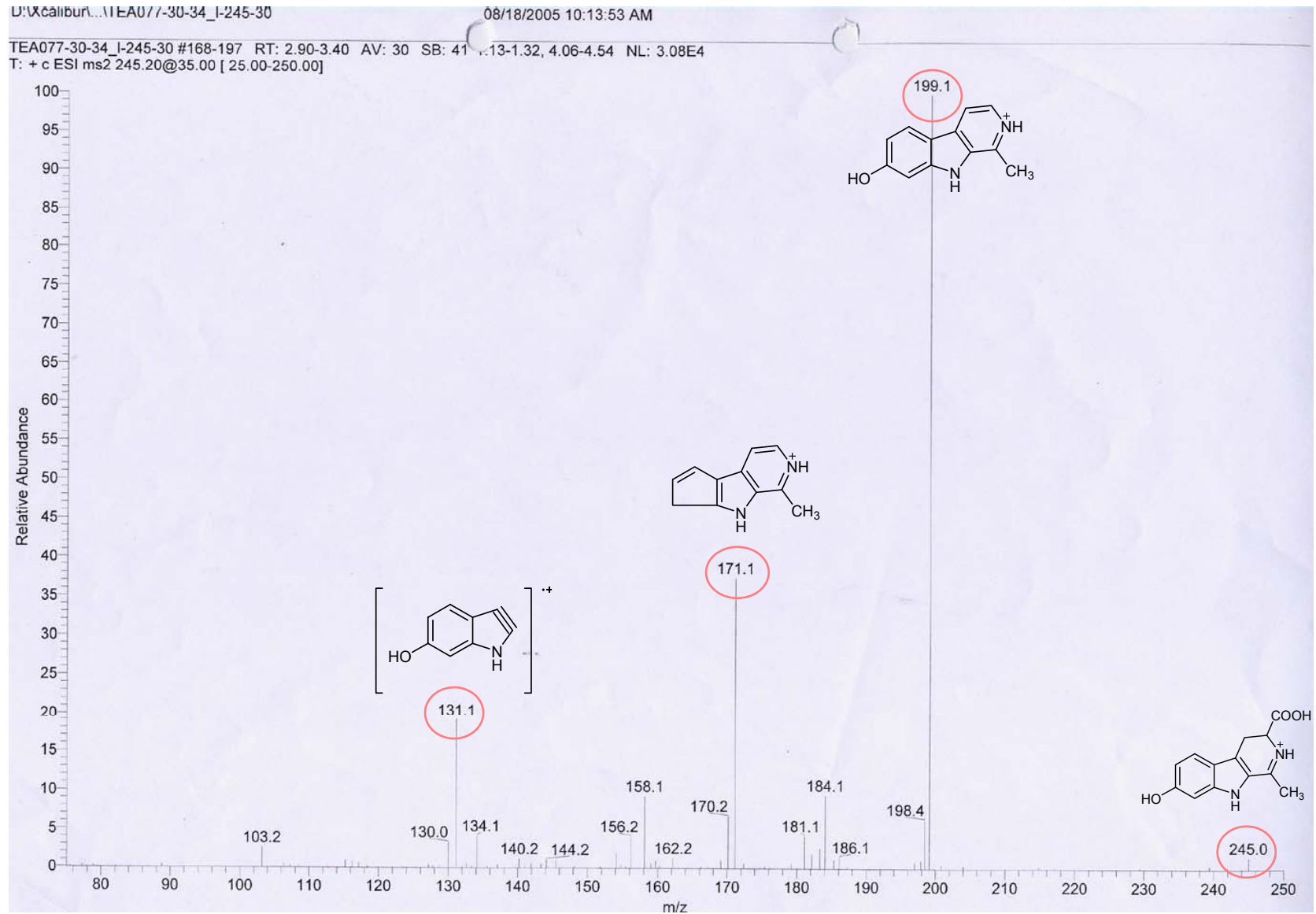
- CD measurement
 - oxidative degradation and GC-MS analysis
- **S-configuration, proposed biosynthetic precursor: L-tryptophane**



Bringmann, G., God, R & Schäfer, M. (1996) – An improved degradation procedure for determination of the absolute configuration in chiral isochinoline and β -carboline derivatives. *Phytochemistry* 43, 1393-1403.

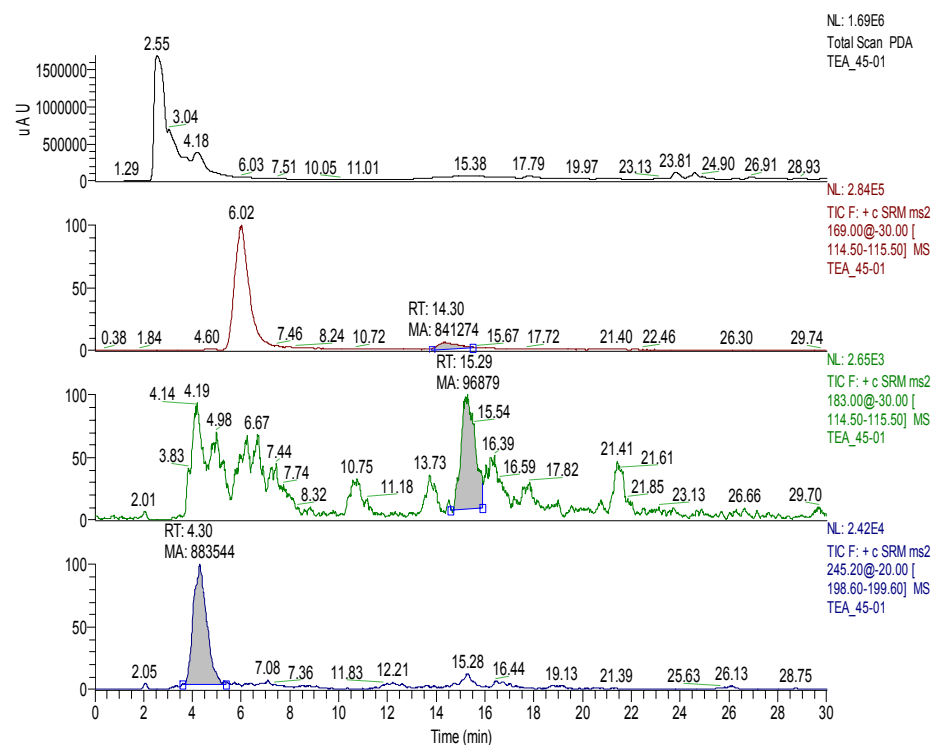
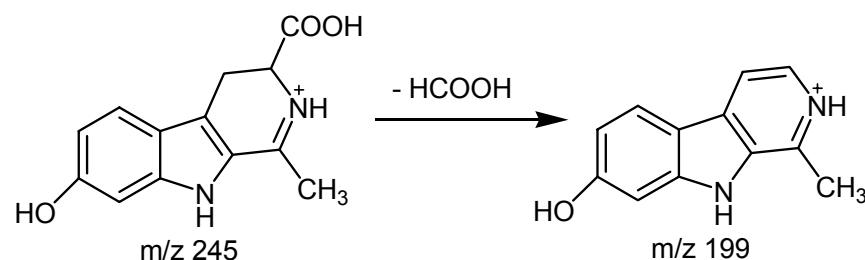


Brunnein A: CID mass spectrum from LC-ESI-MS/MS



Occurrence of Brunnein A in *Hygrophorus* spp.

- SRM measurements:
- Using main CID MS fragmentation of Brunnein A:



Axel Teichert, Å., Schmidt, J., Kuhnt, C., Huth, M., Porzel, A., Wessjohann, L., Arnold, N. (2008) - Determination of β -carboline alkaloids in fruiting bodies of *Hygrophorus* spp. by liquid chromatography/electrospray ionisation tandem mass spectrometry. *Phytochem. Analysis.*, in press.



Occurrence of Brunnein A in *Hygrophorus* spp.

Results:

Brunnein A occurs only in section Olivaceoumbrini within the species *H. pustulatus*, *H. olivaceoalbus*, (*H. personii*), *H. agathosmus*, *H. latitabundus*, *H. hyacinthinus*, (not in *H. marzuolus* !)

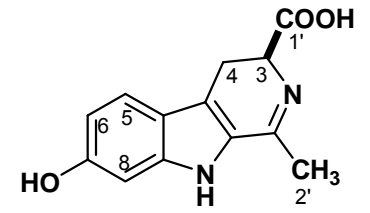
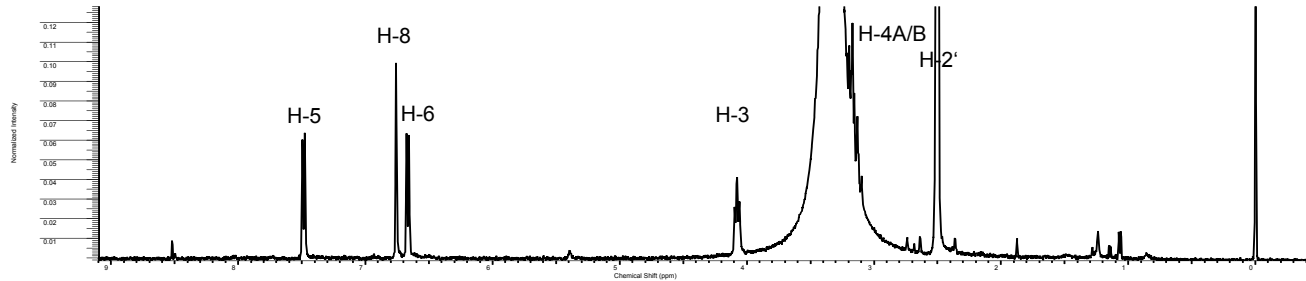
→ chemotaxonomical marker for this section, comparable to taxonomical classification based on morphological data



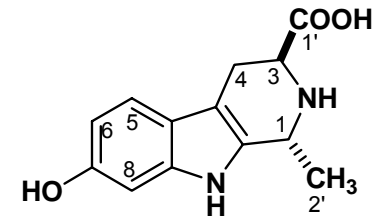
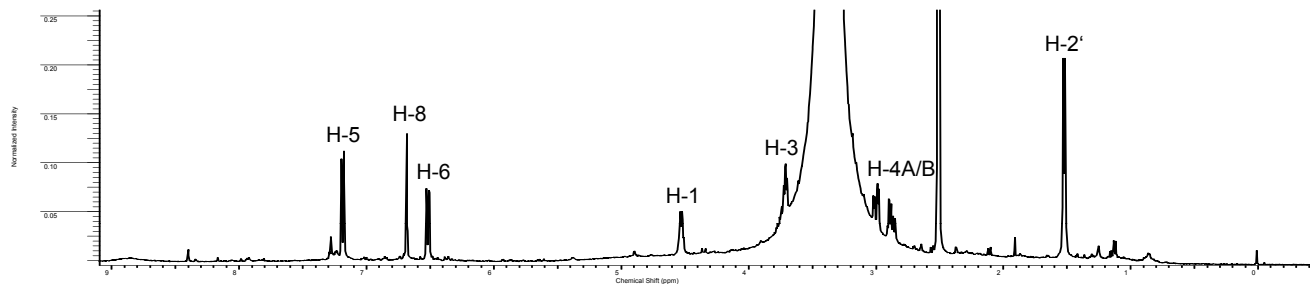
Axel Teichert, Å., Schmidt, J., Kuhnt, C., Huth, M., Porzel, A., Wessjohann, L., Arnold, N. (2008) - Determination of β -carboline alkaloids in fruiting bodies of *Hygrophorus* spp. by liquid chromatography/electrospray ionisation tandem mass spectrometry. *Phytochem. Analysis.*, in press.



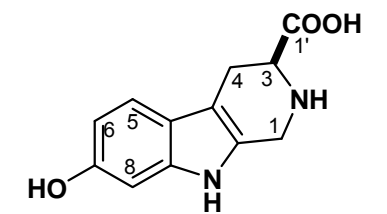
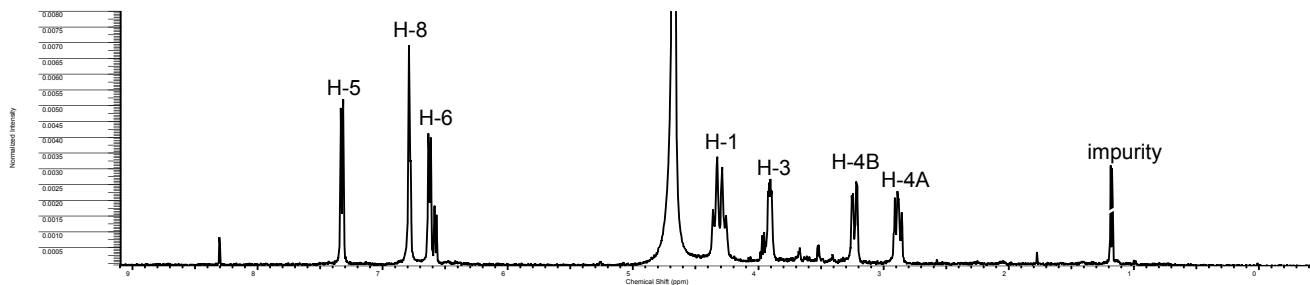
Brunnein A – D: ^1H NMR



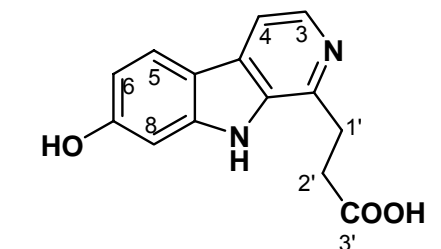
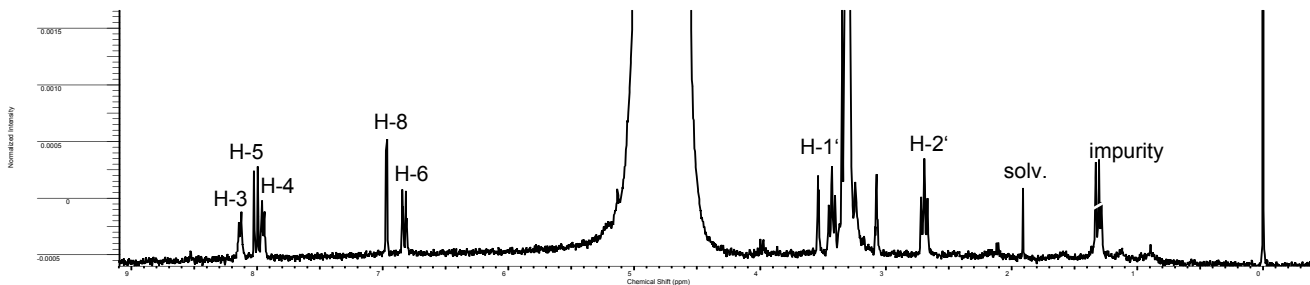
Brunnein A, $\text{DMSO-}d_6$



Brunnein B, $\text{DMSO-}d_6$



Brunnein C, D_2O



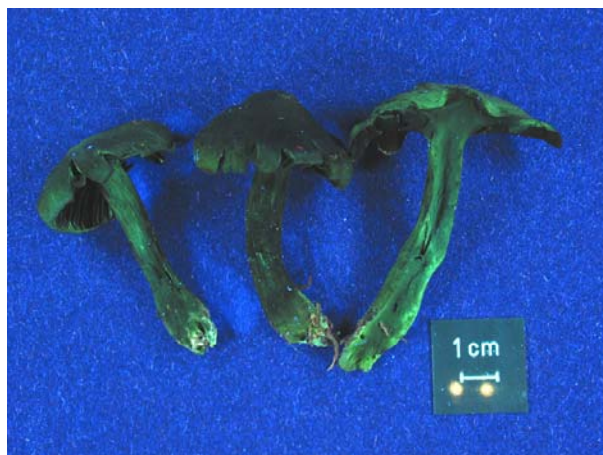
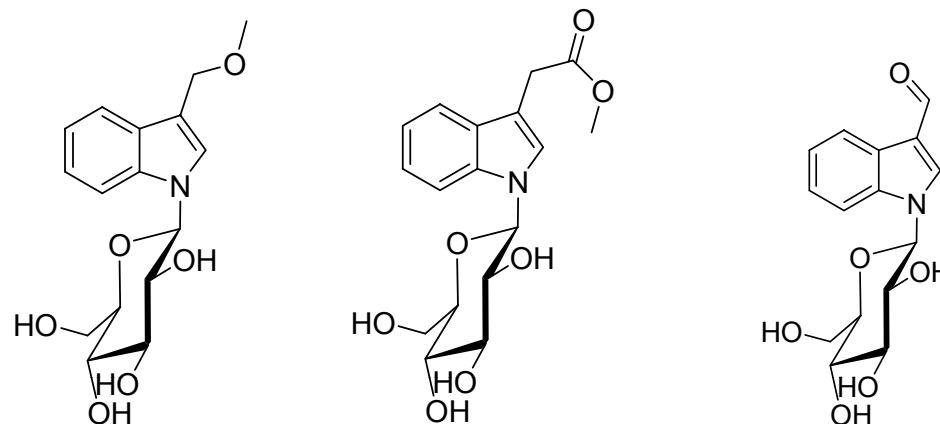
Brunnein D, CD_3OD



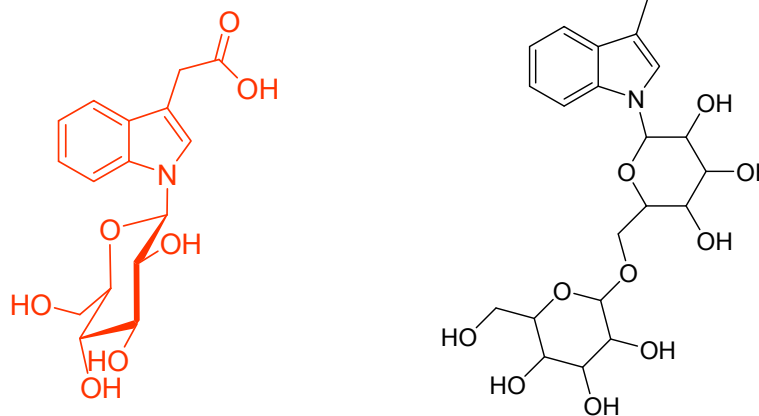
β -Carboline Alkaloids from *Cortinarius brunneus*



Cortinarius brunneus



Cortinarius brunneus (365 nm)



Teichert, A., Schmidt, J., Porzel, A., Arnold, N., Wessjohann, L. (2007) - Brunneins A – C, β -carboline alkaloids from *Cortinarius brunneus* (Basidiomycetes). *J. Nat. Prod.* 70(9), 1529-1531.

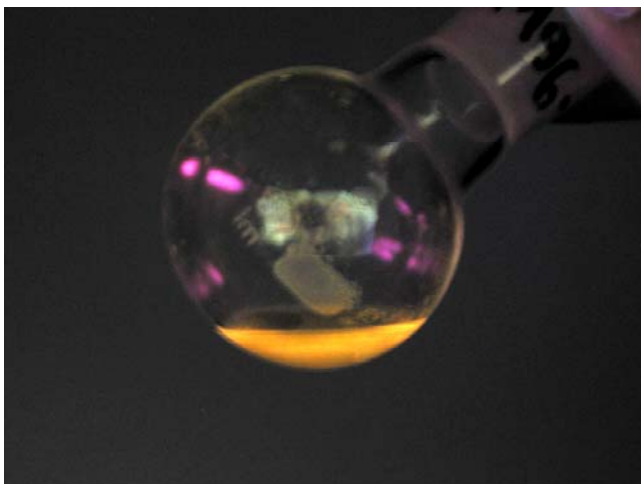
Teichert, A., Schmidt, J., Porzel, A., Arnold, N., Wessjohann, L. (2008) - *N*-Glucosyl Indole Derivatives from *Cortinarius brunneus* (Basidiomycetes). *Chem. Biodiv.* 5, 664-669.



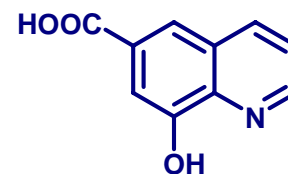
(Iso-) Quinoline Alkaloids from *Cortinarius subtortus*



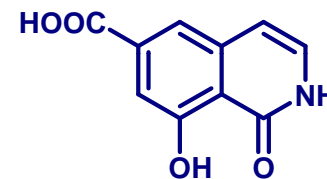
Cortinarius subtortus



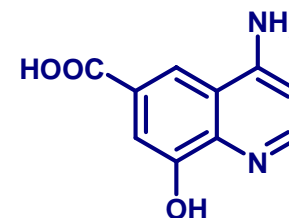
Crude extract



8-Hydroxyquinoline-6-carboxylic acid



8-Hydroxy-1-oxo-dihydroisoquinoline-6-carboxylic acid



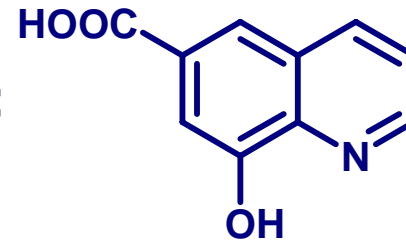
4-Amino-8-hydroxyquinoline-6-carboxylic acid

Teichert, A., Schmidt, J., Porzel, A., Arnold, N., Wessjohann, L. (2008) - (Iso-) Quinoline Alkaloids from *Cortinarius subtortus* *J. Nat. Prod.*, in press.



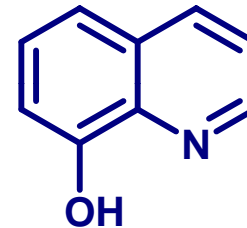
(Iso-) Quinoline alkaloids – similarities!

secondary metabolite from fungi:



8-Hydroxyquinoline-6-carboxylic acid

synthetic compound



8-Hydroxyquinoline

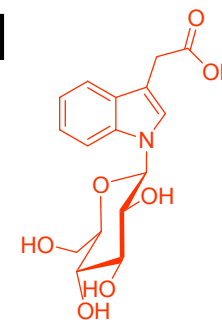
Plant fungicide



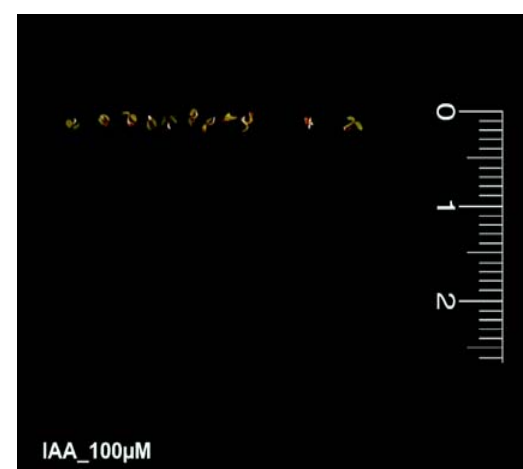
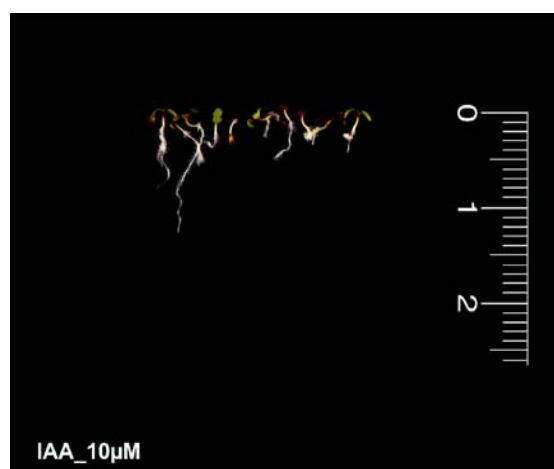
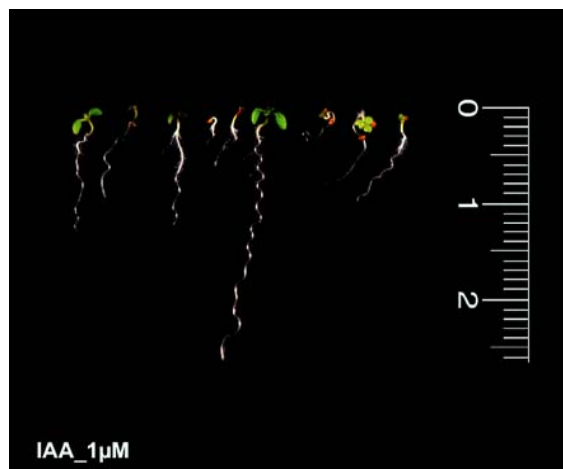
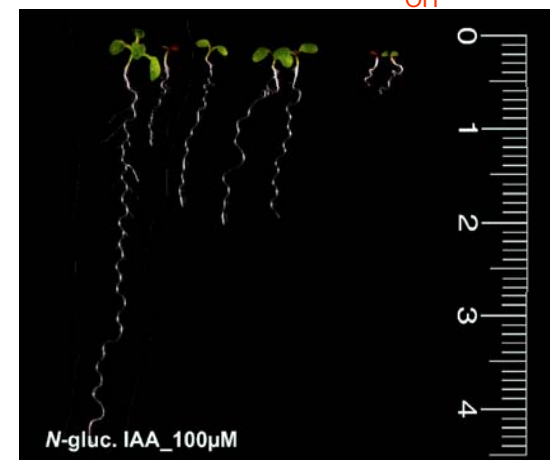
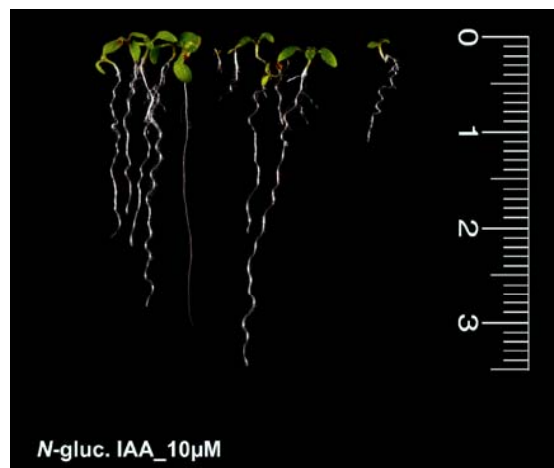
Desinfectants



Bioactivity of *N*-Glucosyl-(1*H*-indol-3-yl)-acetic acid

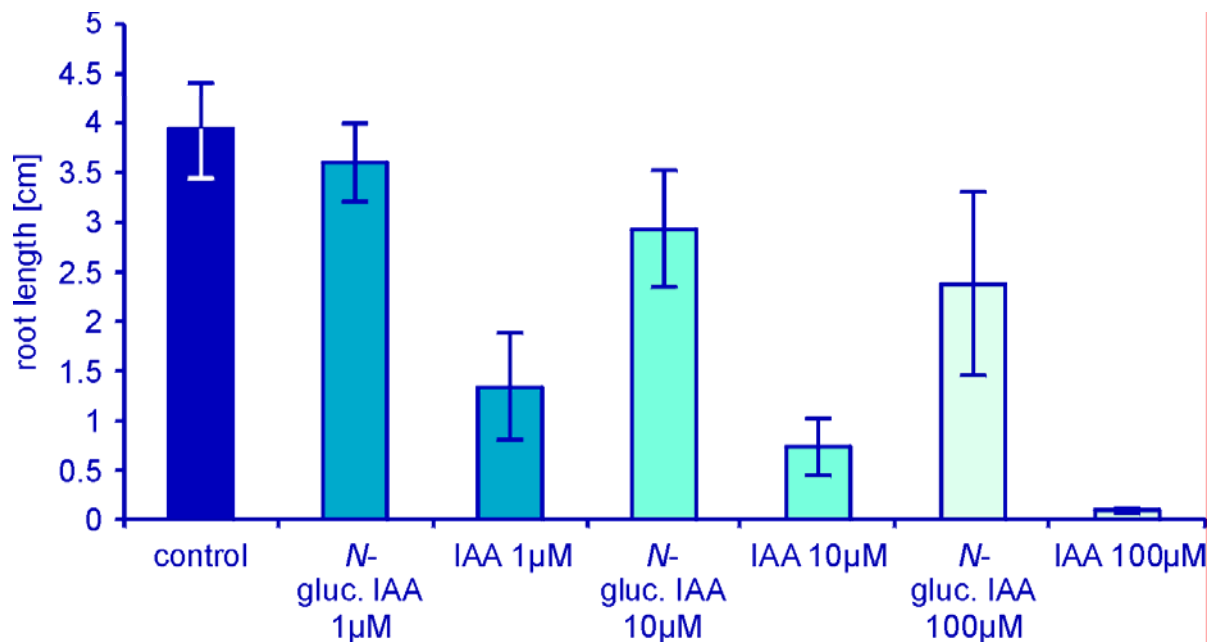
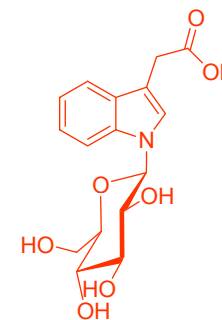


Arabidopsis thaliana tap root elongation assay (10 dpi)



Bioactivity of *N*-Glucosyl-(1*H*-indol-3-yl)-acetic acid

Arabidopsis thaliana tap root elongation assay (10 dpi)



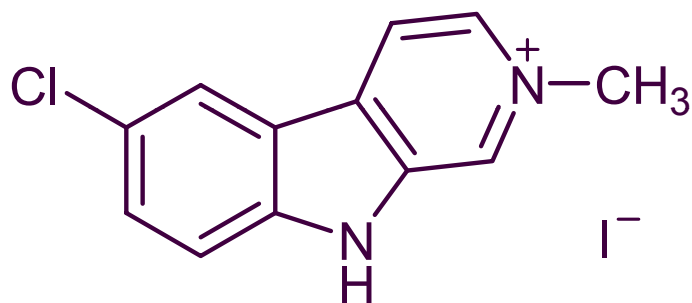
→ no auxin like activity of *N*-gluc IAA

→ possible: inactive storage or transport conjugate?

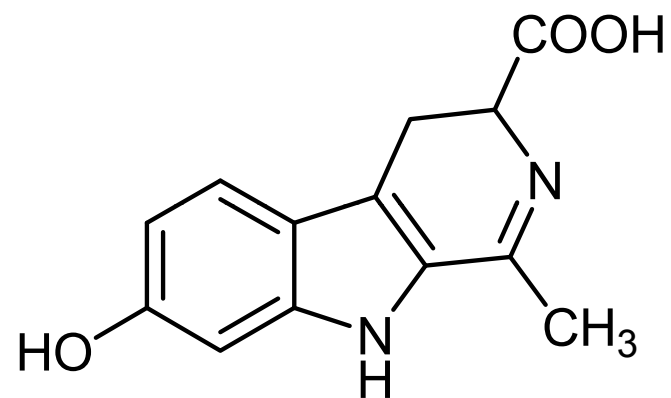


Bioactivity of Brunnein A

structurally similar to nostocarboline¹, a potent AChE inhibitor



Nostocarboline



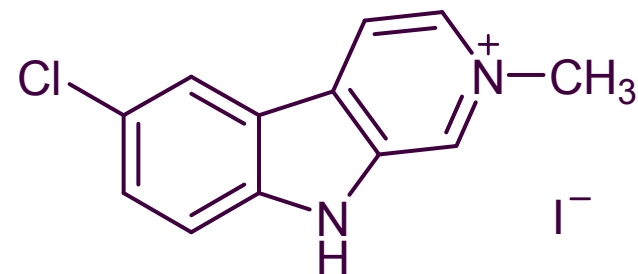
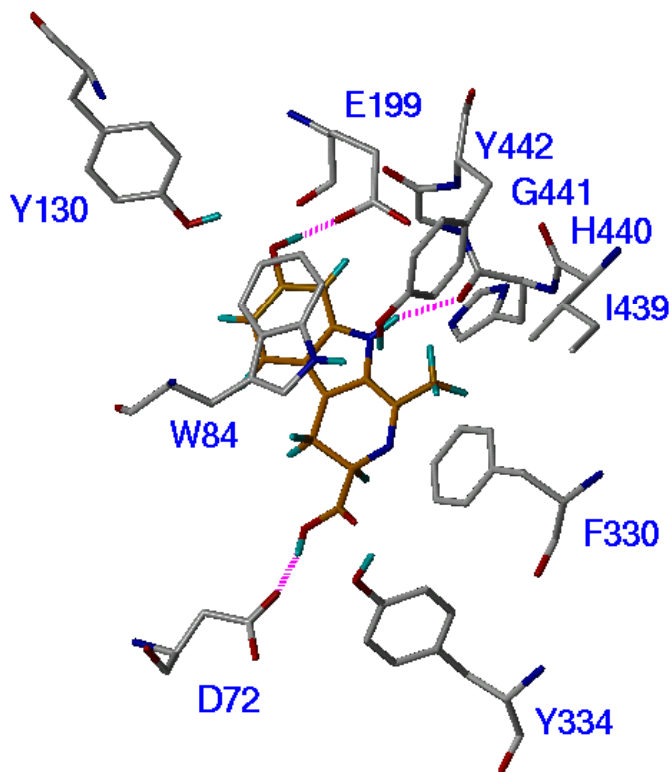
Brunnein A

¹Paul G. Becher, Julien Beuchat, Karl Gademann, Friedrich Jüttner, 2005 - Nostocarboline: Isolation and Synthesis of a New Cholinesterase Inhibitor from *Nostoc* 78-12A, J. Nat. Prod., 68, 1793-1795.

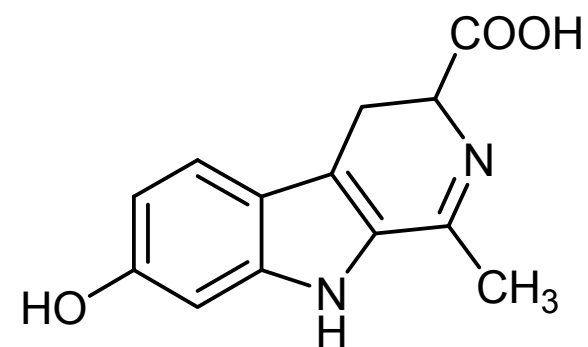


In silico docking studies to AChE

Active site



Nostocarboline

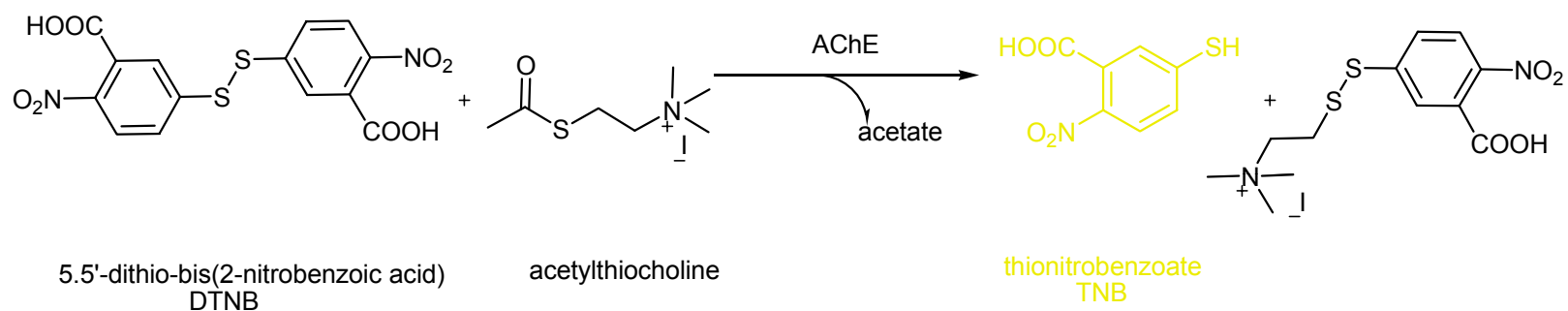


Brunnein A

score of Brunnein A > Nostocarboline: H-H bridge



in vitro AChE inhibition assay using Ellman's method

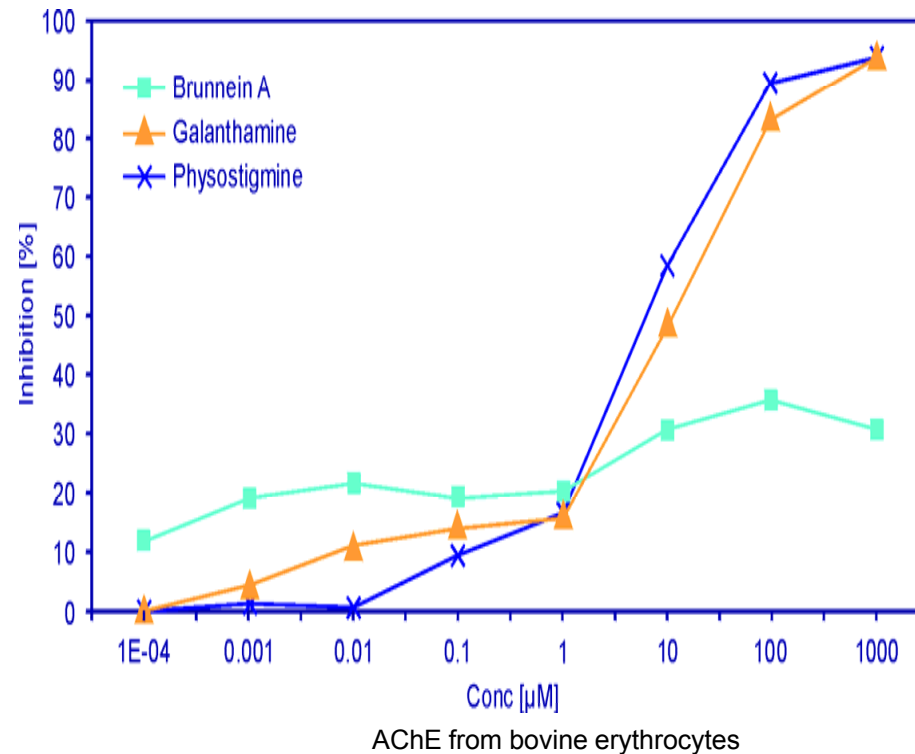


¹ Schott et al., 2006, 6-hydroxy- and 6-methoxy- β -carbolines as acetyl- and butyrylcholinesterase inhibitors, *Bioorg. Med. Chem. Lett.* 12, 5840-5843

² Gearhart et al., 2002, Phenylethanolamine N-methyltransferase has β -carboline 2N-methyltransferase activity..., *Neurochem. Int.* 40, 611-620



in vitro AChE inhibition assay using Ellman's method



- only marginal inhibition effects
- literature: • carbolinium salts best inhibitors, *i.a.* N-Me derivatives¹
 - β -carboline 2N-methyltransferase in human brain²
- pro drug hypothesis \rightarrow tert. carbolines as prodrugs (lipophil - transport)?
 - \rightarrow metabol. in active carbolinium salts (hydrophil - brain locked)?

¹ Schott et al., 2006, 6-hydroxy- and 6-methoxy- β -carboline as acetyl- and butyrylcholinesterase inhibitors, *Bioorg. Med. Chem. Lett.* 12, 5840-5843

² Gearhart et al., 2002, Phenylethanolamine N-methyltransferase has β -carboline 2N-methyltransferase activity..., *Neurochem. Int.* 40, 611-620



Antifungal assay

Cladosporium cucumerinum (scab or gummosis)



Advantage:

- + good initial test system
- + simply and cheap
- + results direct observable



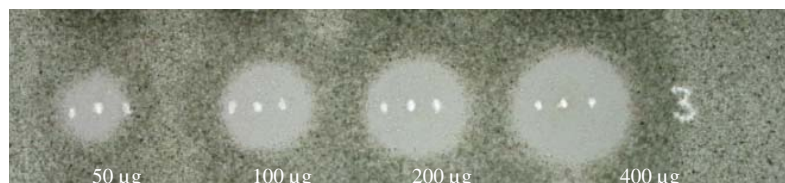
Disadvantage:

- no information about the concentration of the active compounds
- diffusion of the compounds on the TLC layer
- results not or hardly comparable with other fungicidal assays

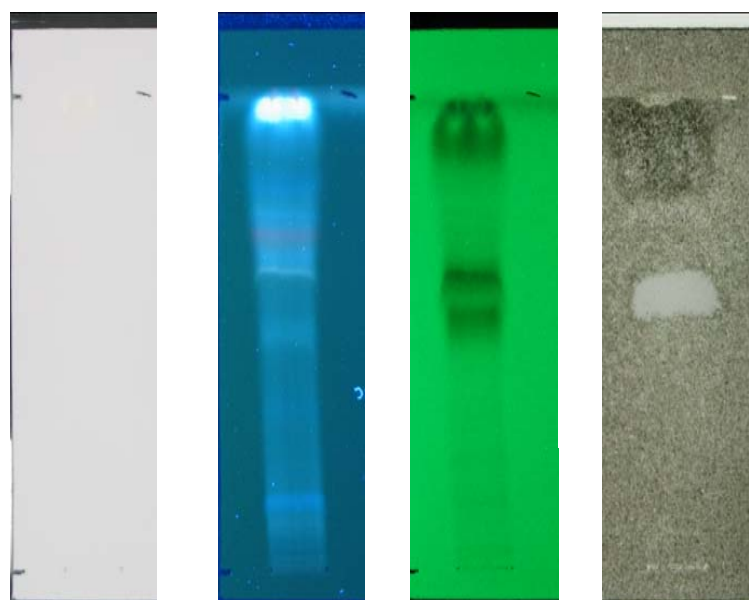
Gottstein, D., Gross, D., Lehmann, H. (1982) – Mikrobiotest mit *Cladosporium cucumerinum* Ell. et Arth. zum Nachweis funitoxischer Verbindungen auf Dünnschichtplatten. Arch. Phytopathol. Pfl. 20, 111 - 116



Direct Bioautography on TLC



EtOAc crude extract *H. eburneus*



daylight

365 nm

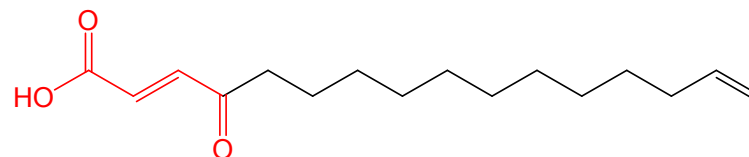
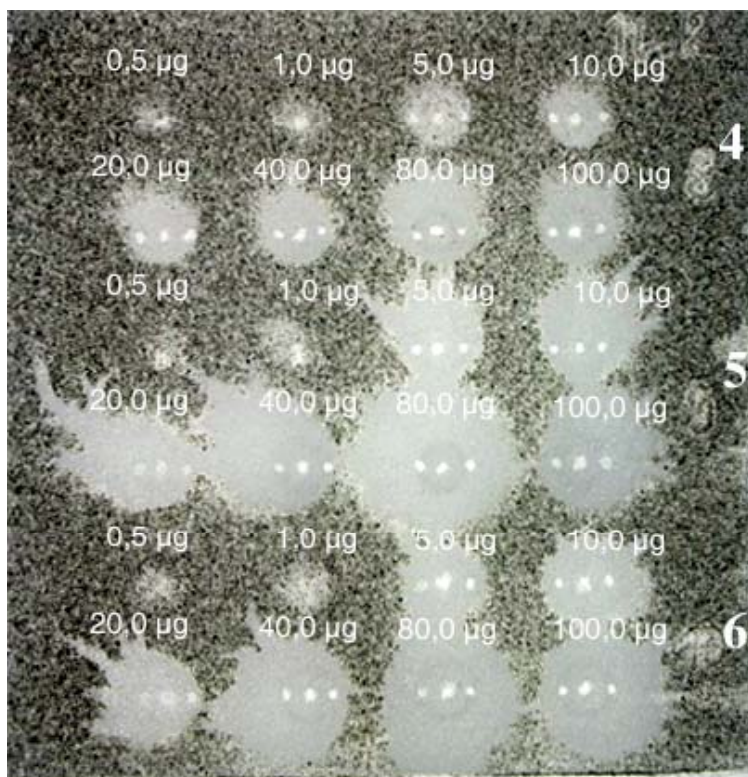
254 nm

incubation

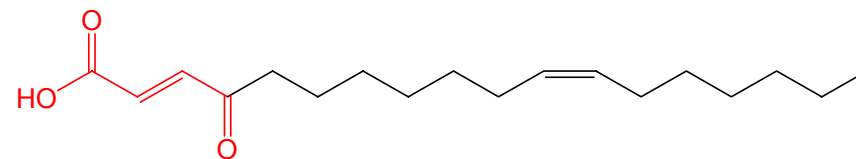
EtOAc crude extract developed on TLC plates



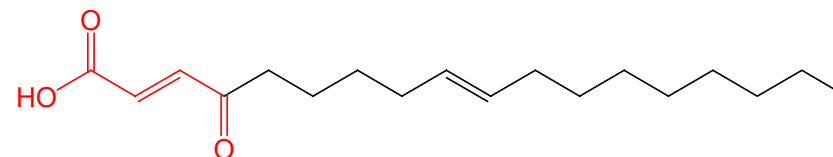
Antifungal activity of 4-Oxo-2-alkenoic fatty acid



(*E*)-4-oxohexadeca-2,15-dienoic acid



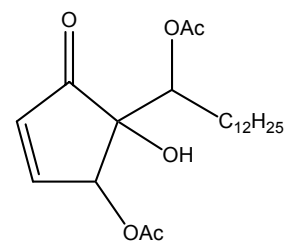
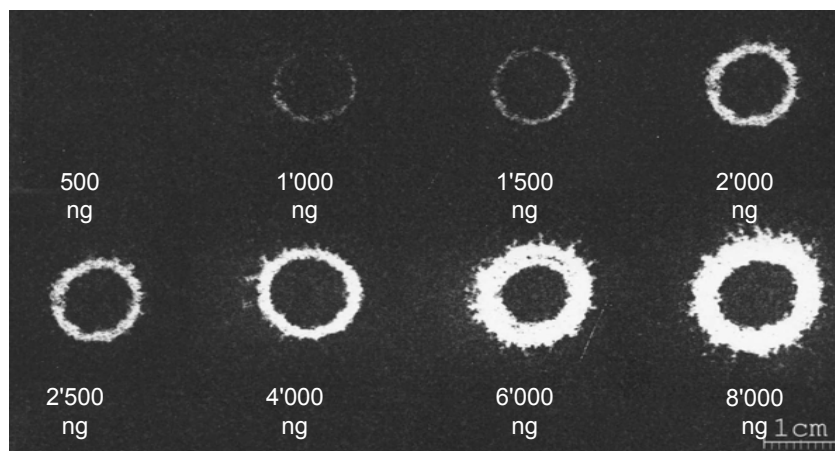
(2*E*,11*Z*)-4-oxooctadeca-2,11-dienoic acid



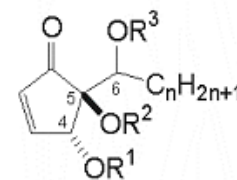
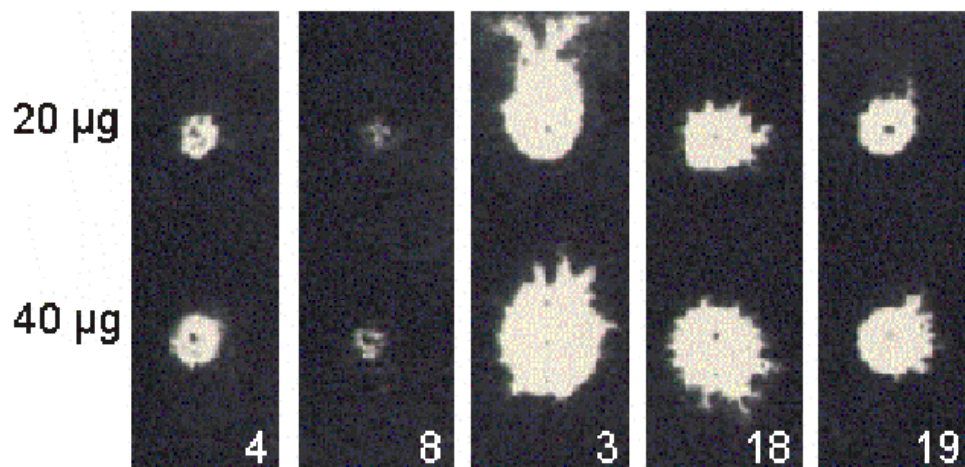
(2*E*,9*E*)-4-oxooctadeca-2,9-dienoic acid



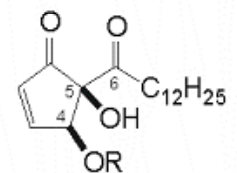
Antifungal activity of Hygrophorones



Hygrophoron A



	R ¹	R ²	R ³	n	
(1)	H	H	H	12	<i>Derivat</i>
(2)	H	H	Ac	12	
(3)	Ac	H	H	12	
(4)	Ac	H	Ac	12	
(5)	Ac	Ac	Ac	12	<i>Derivat</i>
(6)	H	H	Ac	14	
(7)	Ac	H	H	14	
(8)	Ac	H	Ac	14	



	R
(18)	Ac
(19)	H



Antialgal activity

Need for ecologically safe Antifouling agents

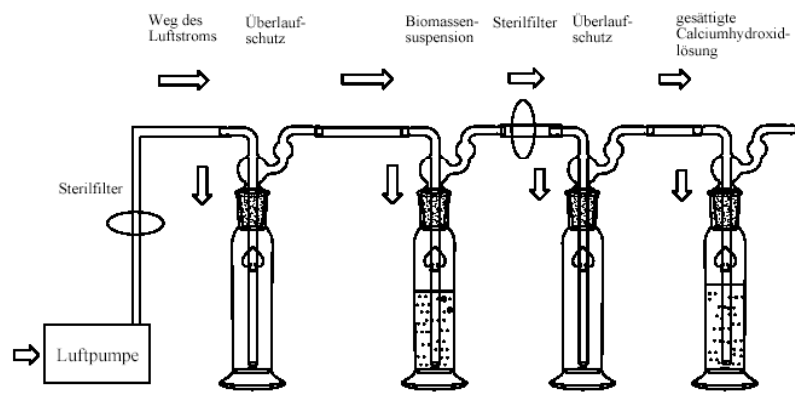


Testorganisms:
Spirulina laxissima
(blue algae)
Scenedesmus rubescens
(green algae)

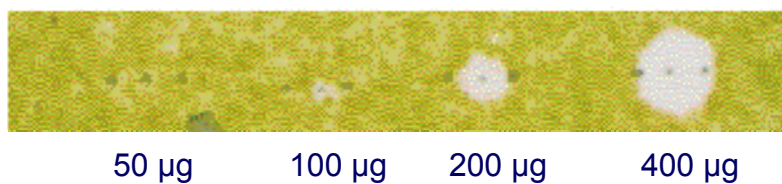


TLC with direct bioautography

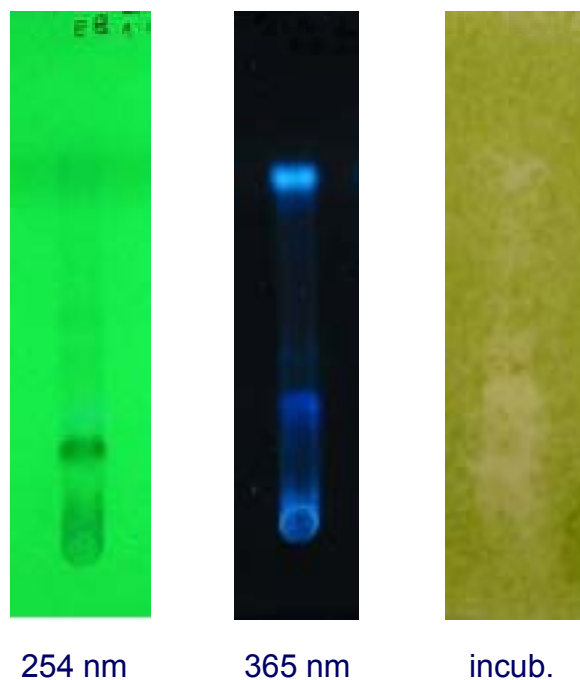
Problem: cultivation and biomass upscale require special equipment and are time consuming:
~ 3-4 weeks upscaling in light chamber and ventilation



Direct Bioautography



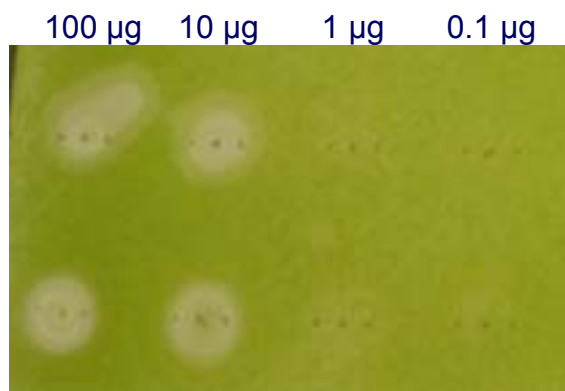
MeOH crude extract *H. eburneus*



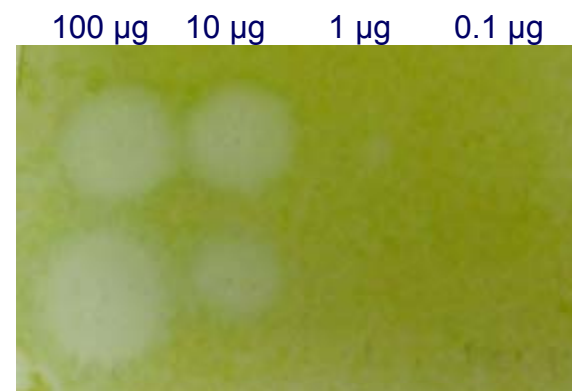
MeOH crude extract developed
on TLC plates



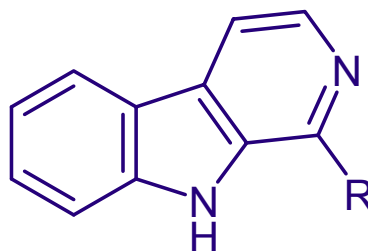
Antialgal activity of Harman and Norharman



Spirulina laxissima



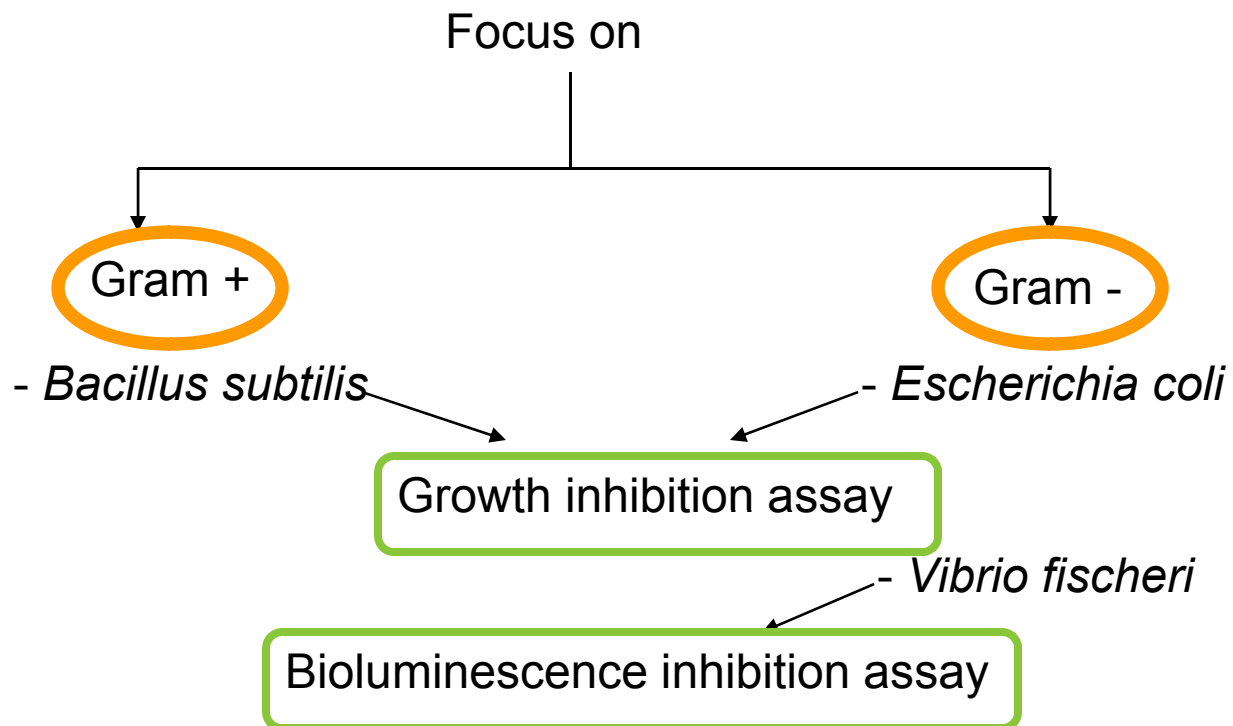
Scenedesmus rubescens



R: H – Norharman
CH₃ – Harman



Antibacterial bioassays



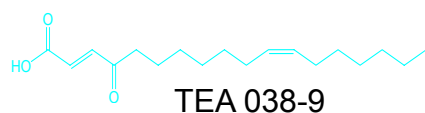
Antibacterial activity: Growth inhibition assay of 4-Oxo-2-alkenoic fatty acid

E. coli (-)

Streptomycin

Amikacin

Rifampicin



100

10

1

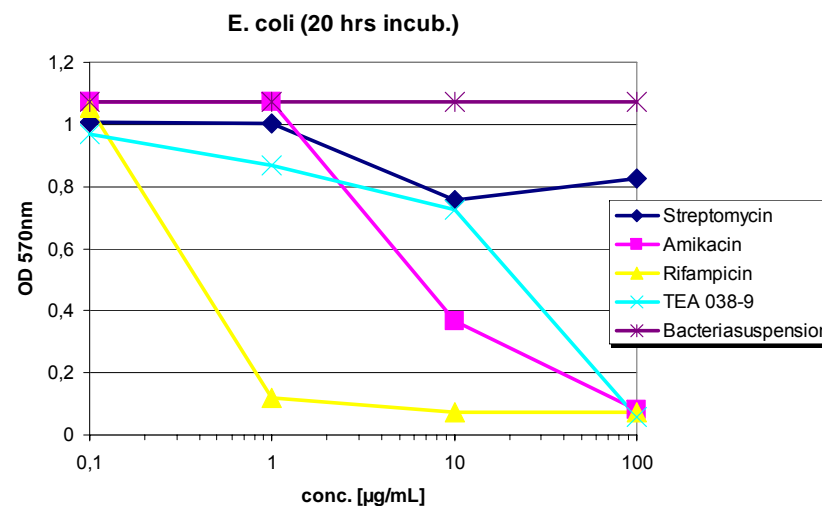
0,1 µg/mL

a) visual evaluation

	100	10	1	0,1 g/mL
Streptomycin	-	-	-	-
Amikacin	++	+	-	-
Rifampicin	++	++	++	-
TEA 038-9	++	+	-	-

++ : strong inhibition; + : weak inhibition; - : no inhibition

b) optical evaluation



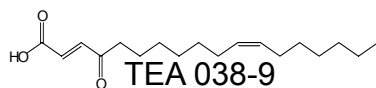
Antibacterial activity: Growth inhibition assay of 4-Oxo-2-alkenoic fatty acid

B. subtilis (+)

Streptomycin

Amikacin

Rifampicin



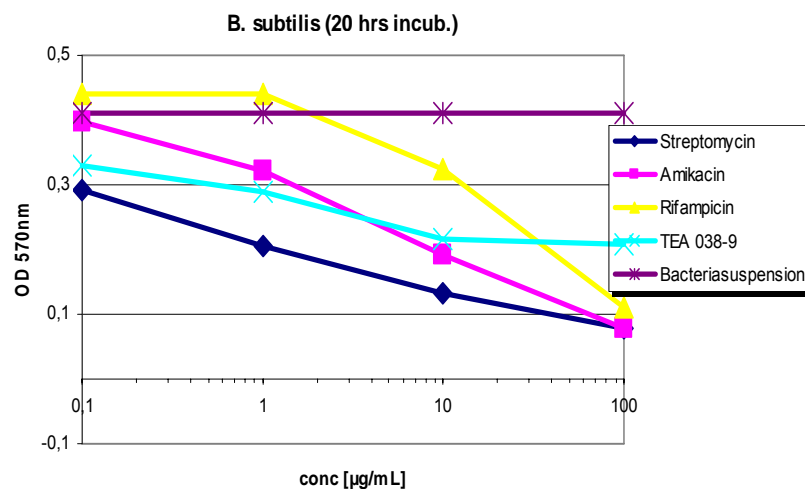
100 10 1 0,1 µg/mL

a) visual evaluation

	100	10	1	0,1 g/ml
Streptomycin	++	+	-	-
Amikacin	++	+	-	-
Rifampicin	++	+	-	-
TEA 038-9	+	+	-	-

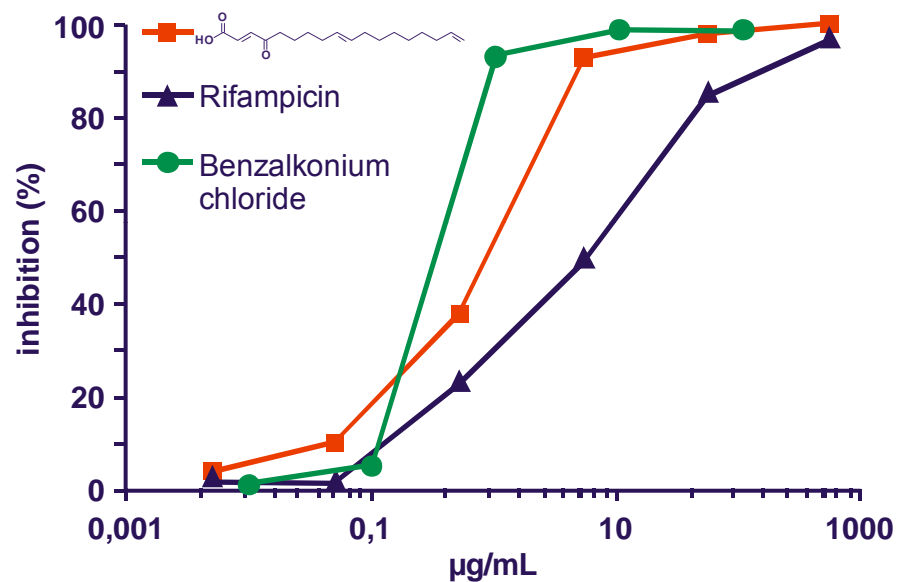
++ : strong inhibition; + : weak inhibition; - : no inhibition

b) optical evaluation



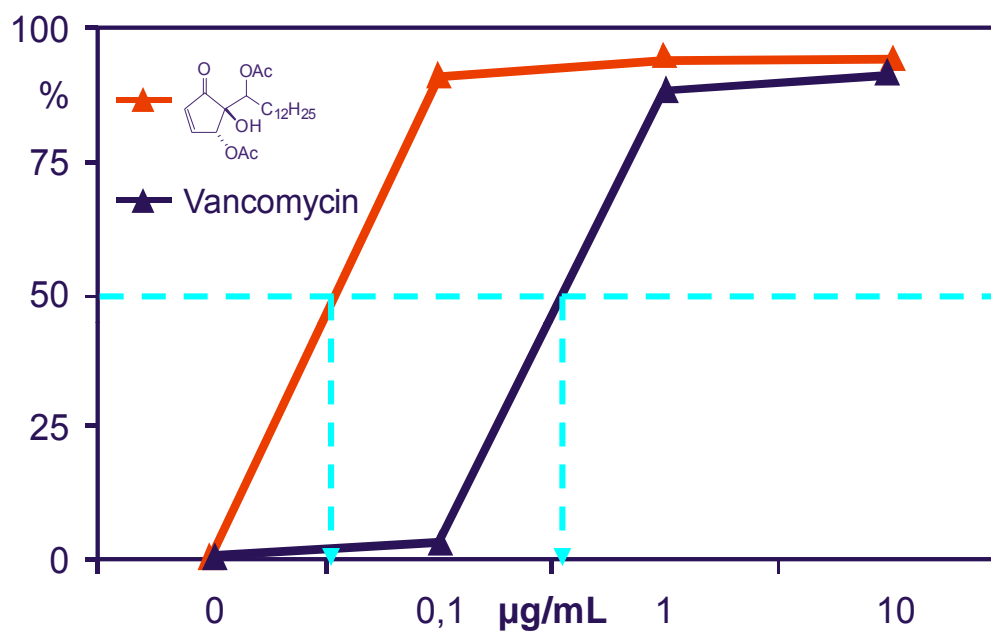
Antibacterial activity: Bioluminescence inhibition assay of 4-Oxo-2-alkenoic fatty acid

Vibrio fischeri (-)



Antibacterial activity: Growth inhibition assay of selected Hygrophorones

methicillin resistant *Staphylococcus aureus* (+)



Anti-Oomycete activity

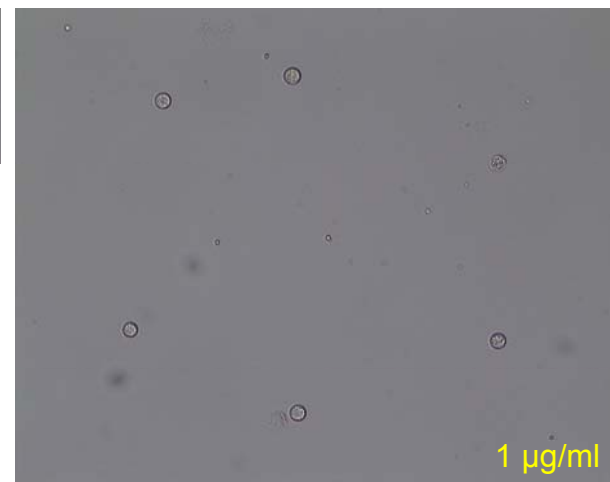
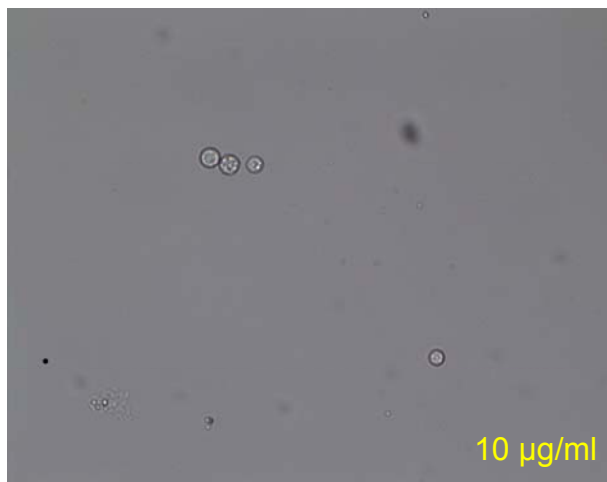
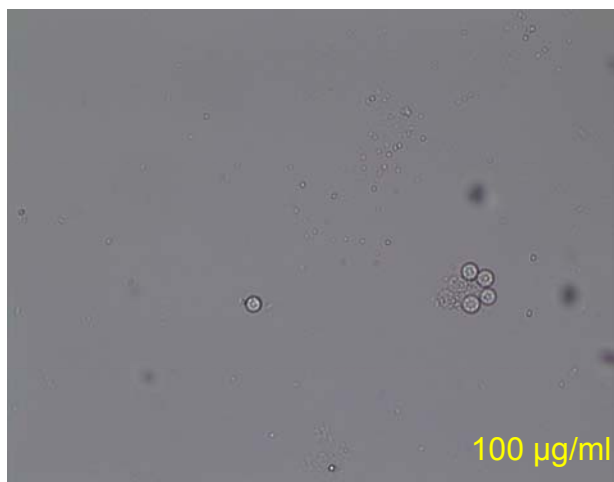
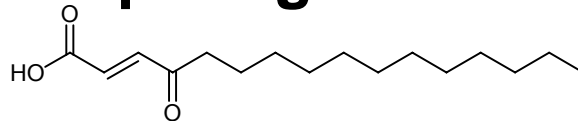


P. infestans

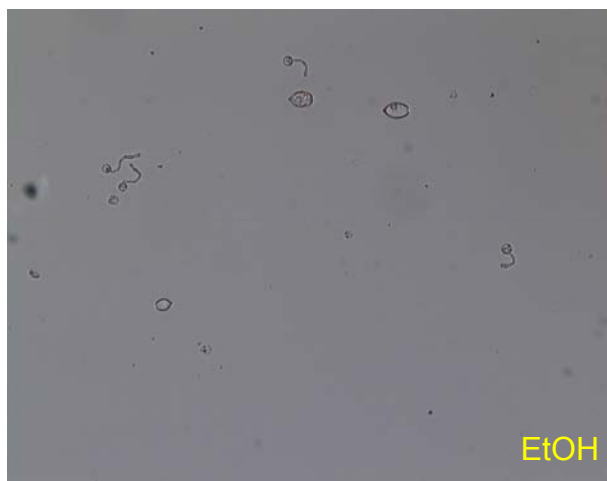
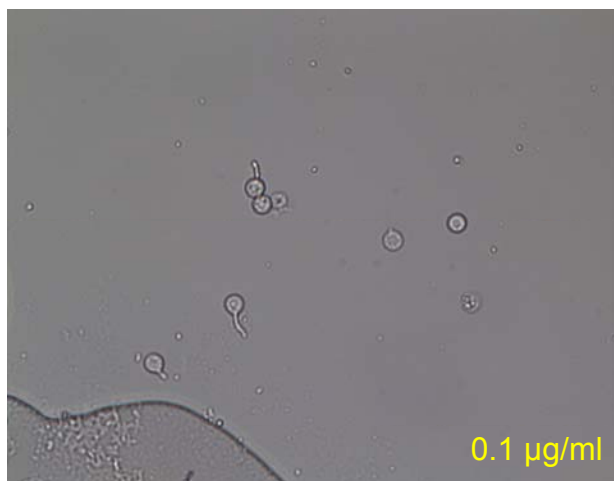
- pathogen of potato blight (Irish potato famine 1845-1850)
- 2 different bioassays:
 - a) spore germination inhibition assay: evaluation under microscope
 - b) mycelial growth inhibition assay: evaluation via fluorescence measurements using a strain expressing GFP as a vital marker



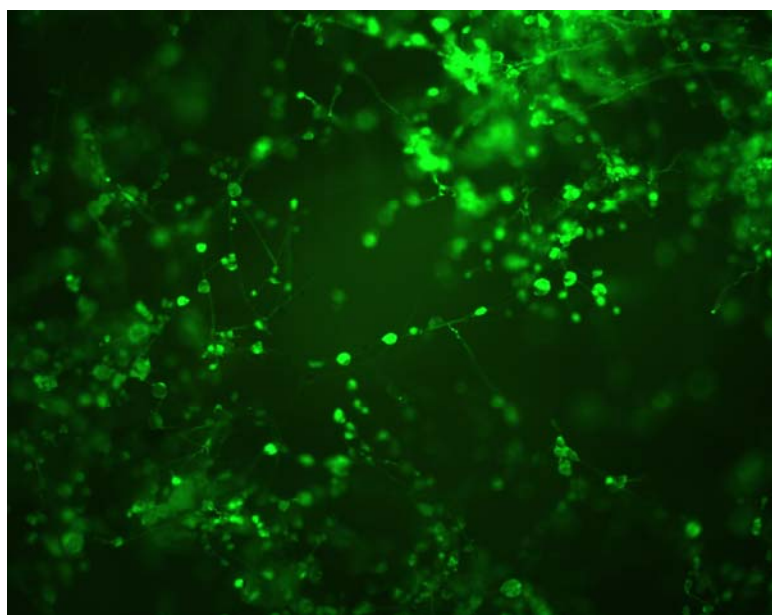
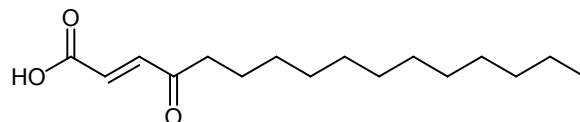
P. infestans spore germination assay



Many bursted spores



P. infestans mycel growth inhibition assay



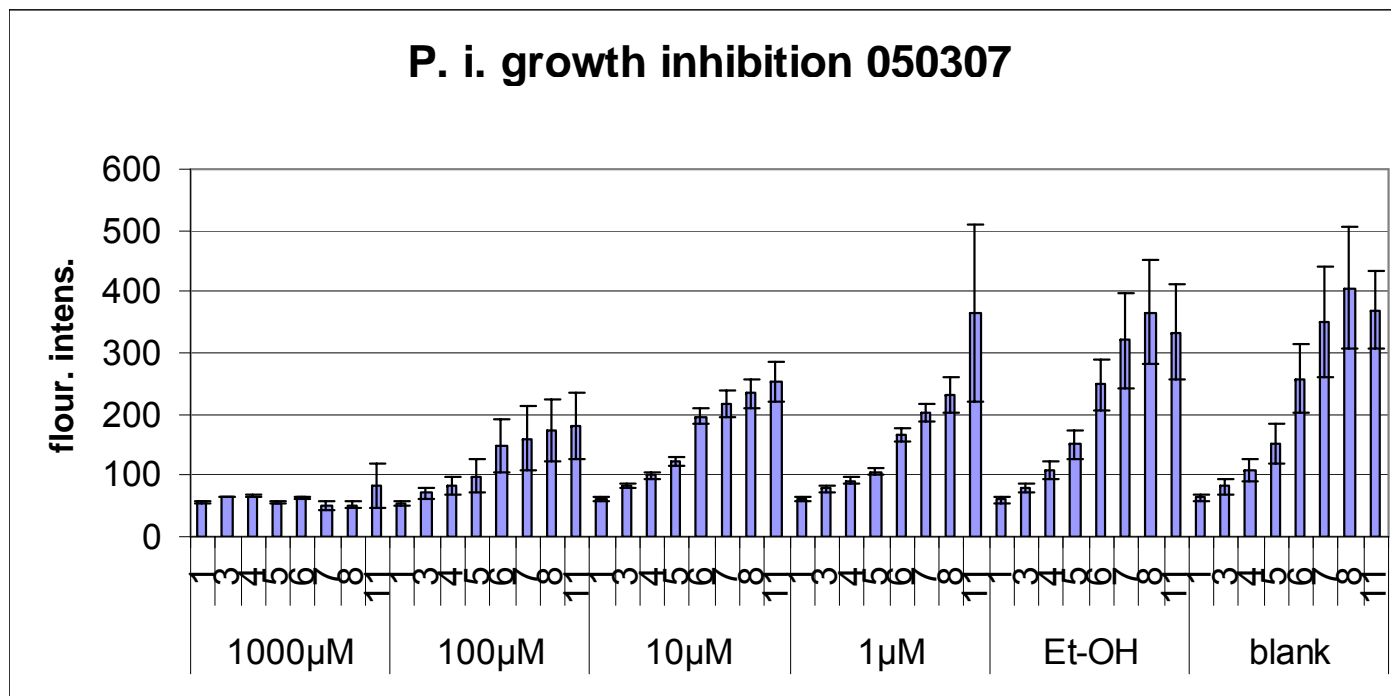
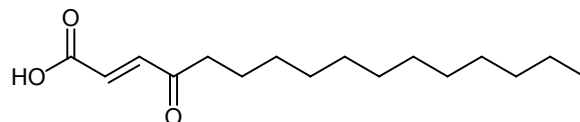
GFP labeled *P. infestans* strain



Visual evaluation after 11 dpi



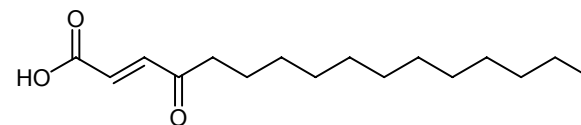
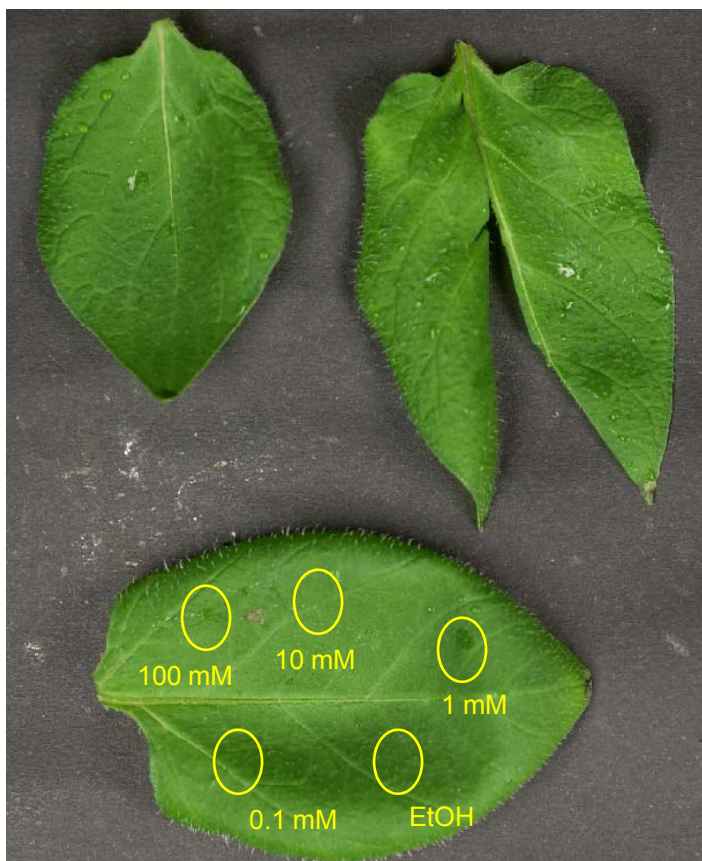
P. infestans mycel growth inhibition assay



Fluorescence intensity measurements after 11 dpi



P. infestans mycel growth inhibition assay: toxicity for *Solanum tuberosum* leaves



3 dpi, 4 different concentrations

No necrotic effects visible



Conclusion

Secondary metabolites from *Hygrophorus* (and *Cortinarius* ?) act as chemical defense substances

Bacterizidal effects

Hygrophorones: very potent against (+) gram bacteria (MRSA)

Fatty acids: high activity against (–) gram bacteria (*V. fischeri*, *E. coli*)

Fungizidal effects

Hygrophorones: high activity against *C. cucumerinum*

Fatty acids: high activity against *C. cucumerinum*

Oomycetes

Fatty acids: high activity in spore germination and mycel growing assay (*P. infestans*)

Algizidal effects

Harman and Norharman: active against *Sp. laxissima*, *Sc. Rubescens*

AChE-assay

Brunnein A: weak activity



Co - worker

Axel Teichert

Monika Kummer

Dr. Tilo Lübken

Dr. Jürgen Schmidt

Dr. Andrea Porzel

Prof. Dr. L. Wessjohann, H



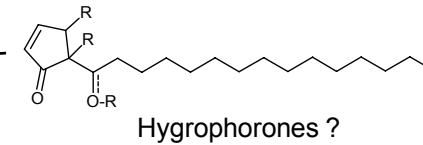
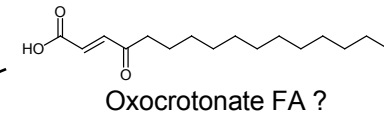
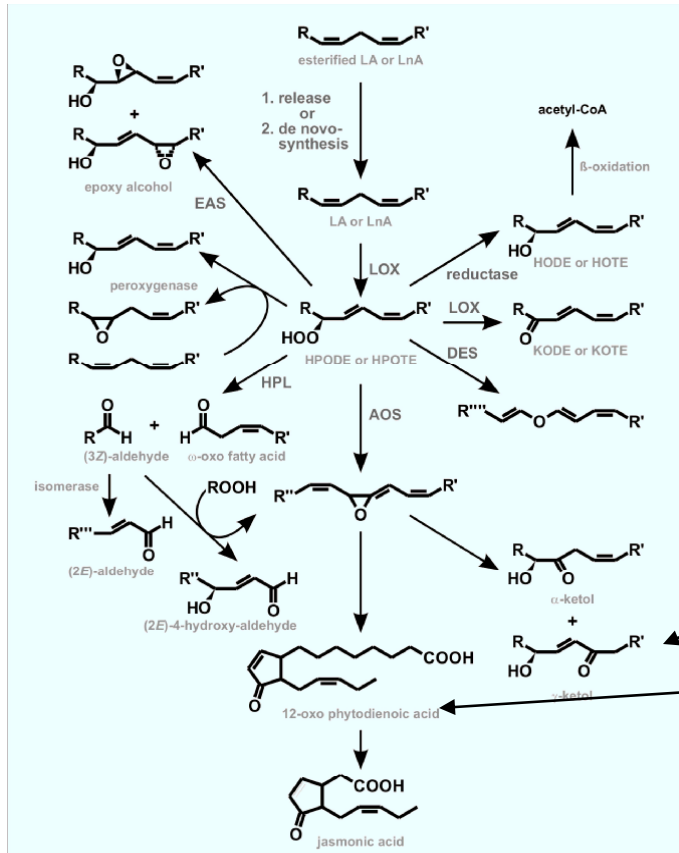
Financial support

DFG, MK LSA



Plant hormones – pathogen defense

→ Similarities in *Hygrophorus* ?!

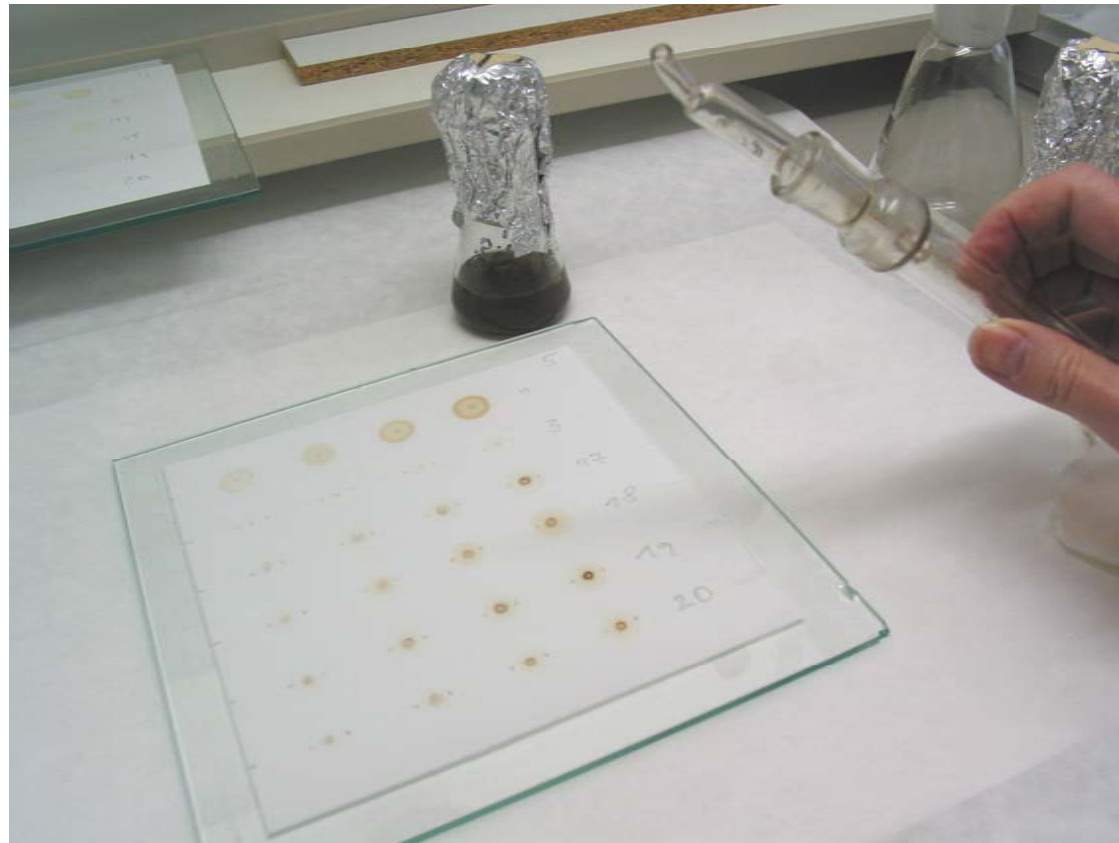


Fungi system ?

Plant system –
jasmonic acid pathway induced by pathogens
including lots of antimicrobial oxylipins



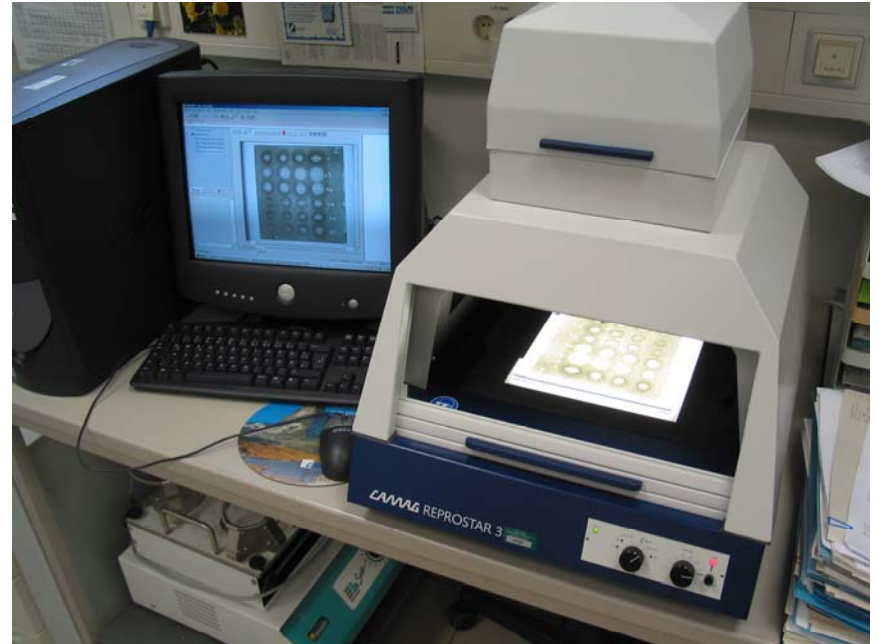
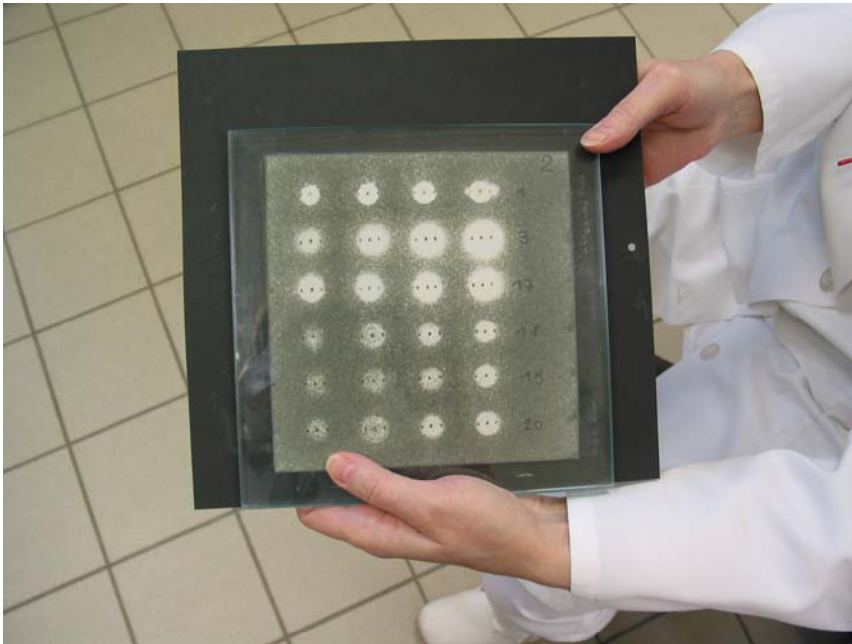
Spraying of the TLC plates with conidiospores of *C. cucumerinum*



Growing of the coloured mycelia in a moisty atmosphere (2 days, 25°C)

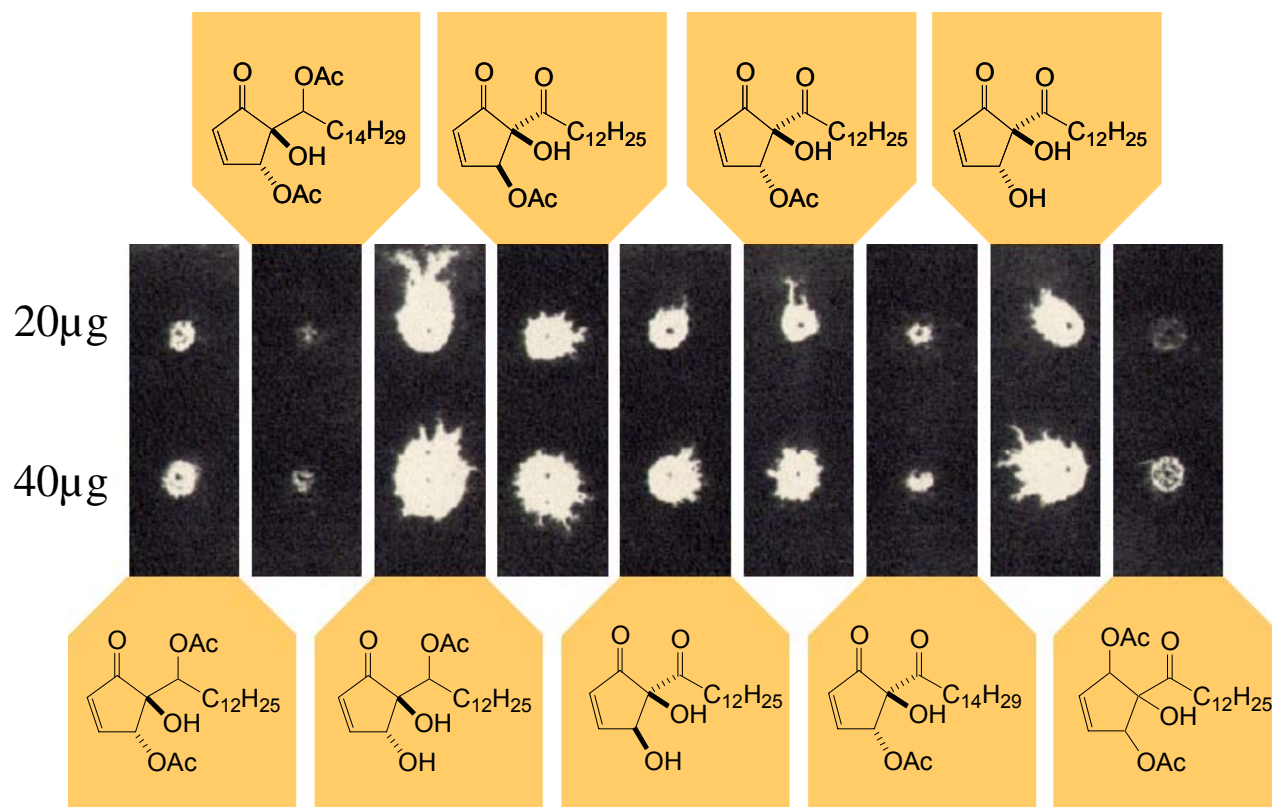


Dokumentation



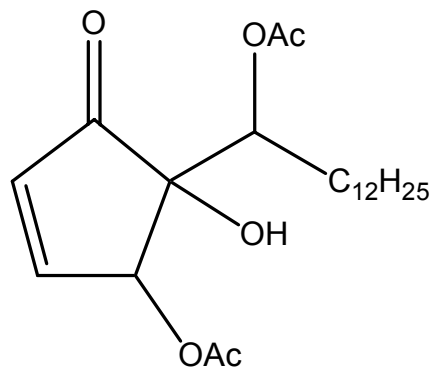
Bioactivity of Hygrophorones A – G

fungicidal activity: *C. cucumerinum*

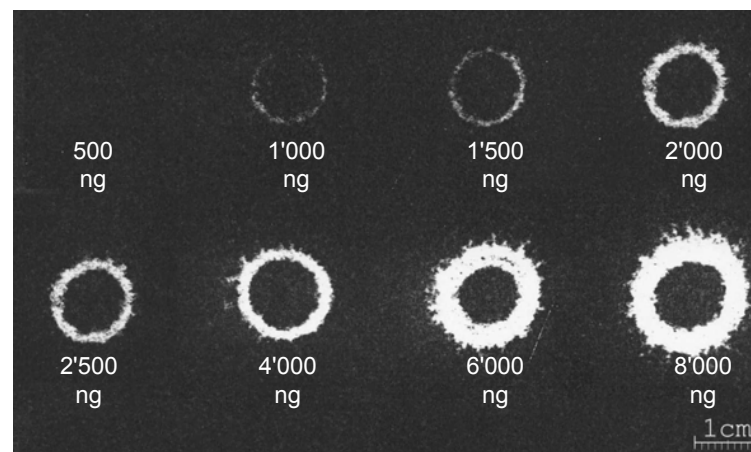


Bioactivity of Hygrophorones

fungicidal activity: *C. cucumerinum*

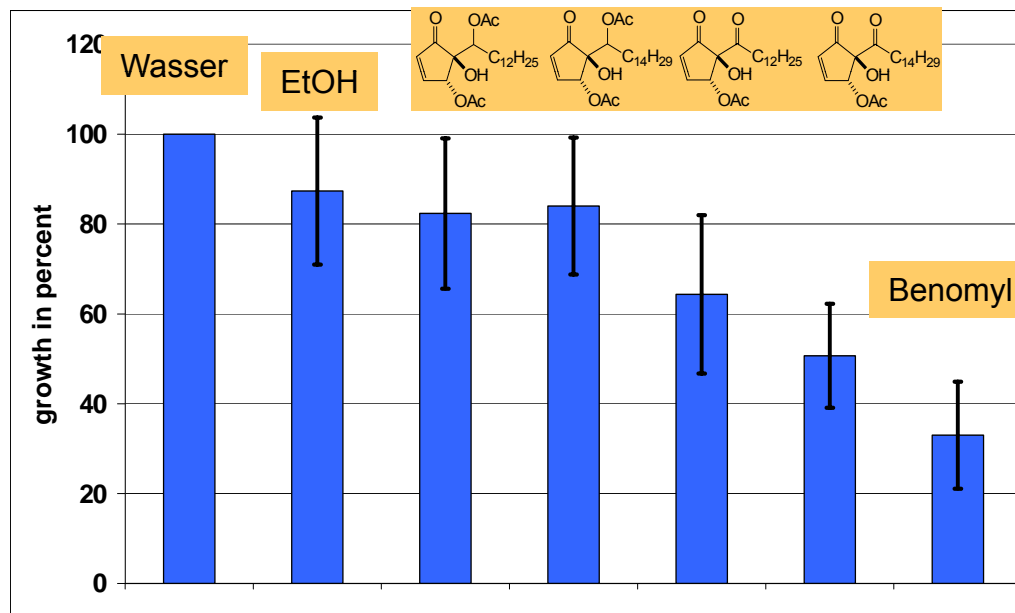


Hygrophoron A



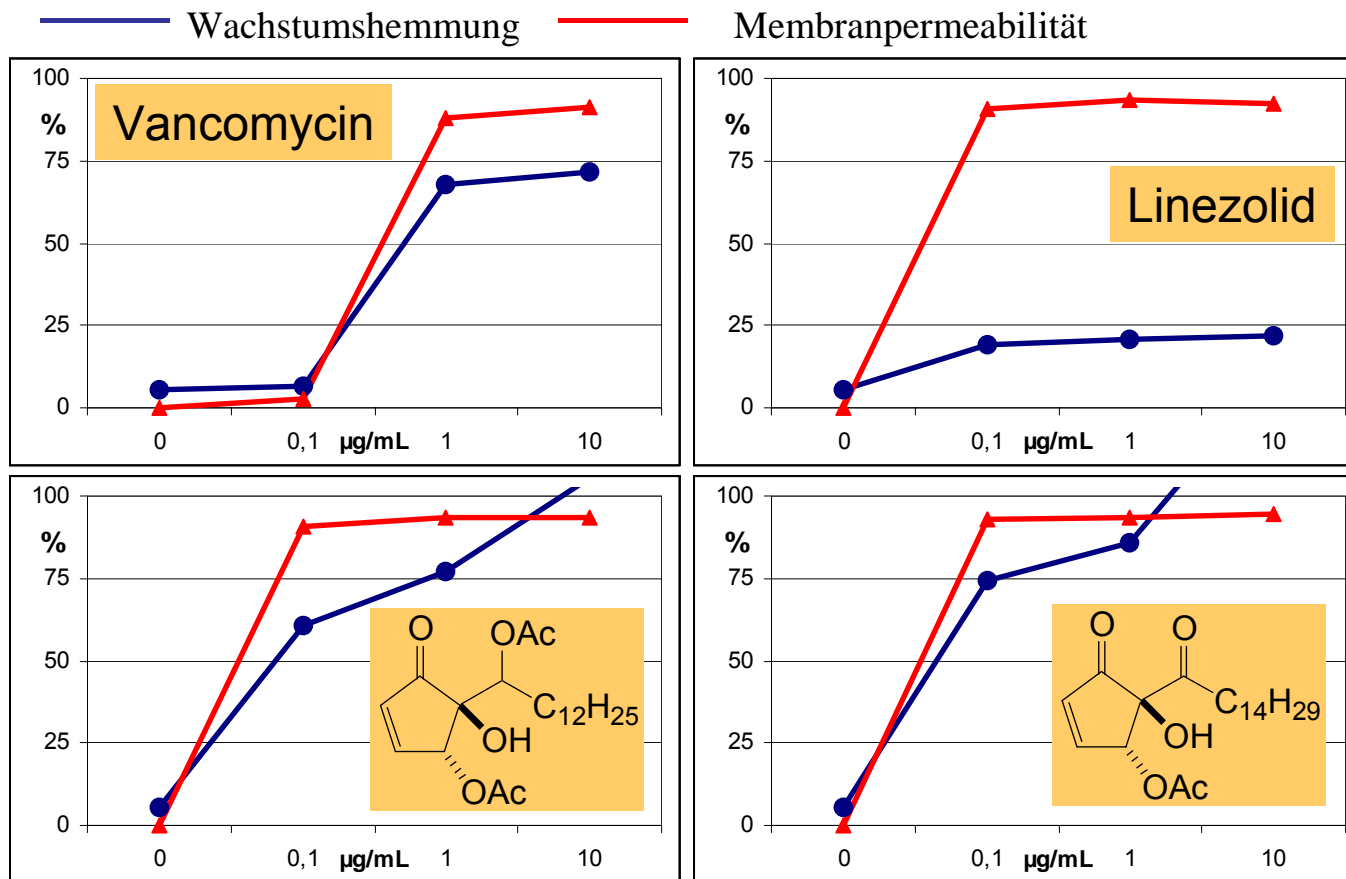
Bioactivity of Hygrophorones

fungicidal activity: *Phytophthora infestans* responsible for potato light



Bioactivity of Hygrophorones

bactericidal activity: MSR *Staphylococcus aureus*



Hygrophorus Sect. Hygrophorus



H. cossus



H. eburneus



H. discoxanthus

<i>species</i>	<i>host</i>	<i>colour in exsiccate</i>
<i>H. cossus</i>	<i>Quercus</i>	white to ochraceous
<i>H. discoxanthus</i>	<i>Fagus</i>	brown to dark brown
<i>H. eburneus</i>	<i>Fagus</i>	white to ochraceous
<i>H. picea</i>	<i>Picea</i>	white to ochraceous

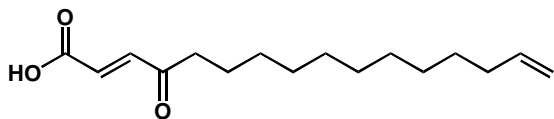


Cossus cossus L.

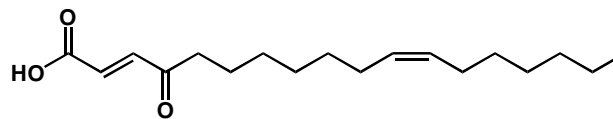
(= Goat Moth, from the strong 'goaty' odour of the caterpillar)



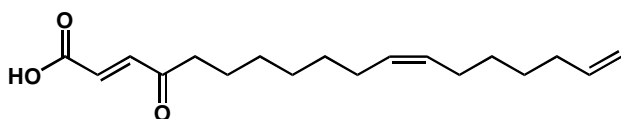
Unusual bioactive fatty acids from *H. eburneus*



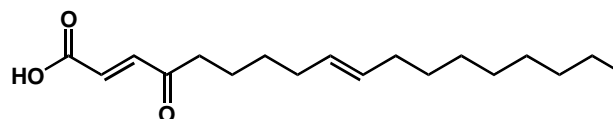
(*E*)-4-oxohexadeca-2,15-dienoic acid (1)



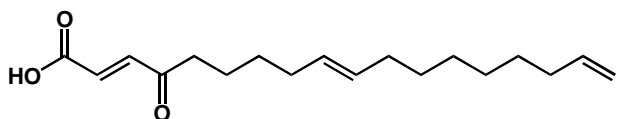
(2*E*,11*Z*)-4-oxooctadeca-2,11-dienoic acid (5)



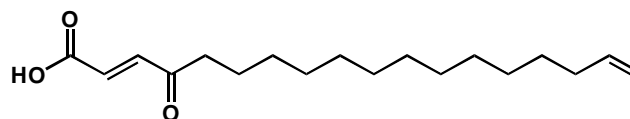
(2*E*,11*Z*)-4-oxooctadeca-2,11,17-trienoic acid (2)



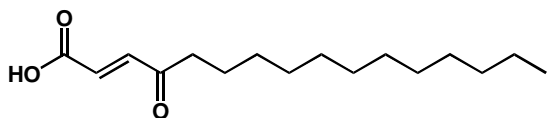
(2*E*,9*E*)-4-oxooctadeca-2,9-dienoic acid (6)



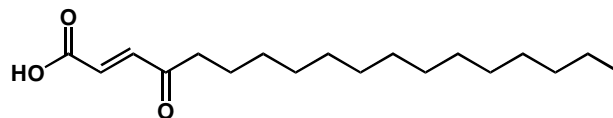
(2*E*,9*E*)-4-oxooctadeca-2,9,17-trienoic acid (3)



(*E*)-4-oxooctadeca-2,17-dienoic acid (7)



(*E*)-4-oxohexadeca-2-enoic acid (4)

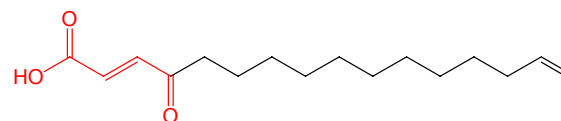
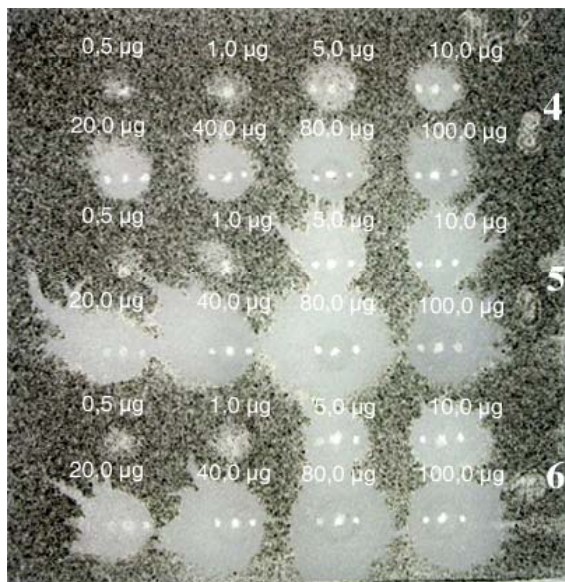


(*E*)-4-oxooctadeca-2-enoic acid (8)

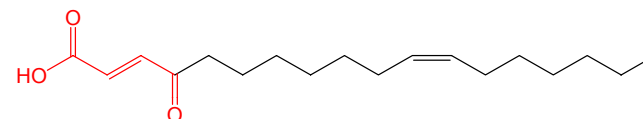


Bioactivity of the isolated fatty acids

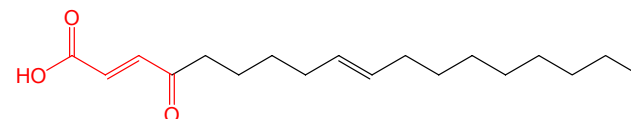
fungicidal activity: *C. cucumerinum*



(*E*)-4-oxohexadeca-2,15-dienoic acid



(*2E,11Z*)-4-oxooctadeca-2,11-dienoic acid

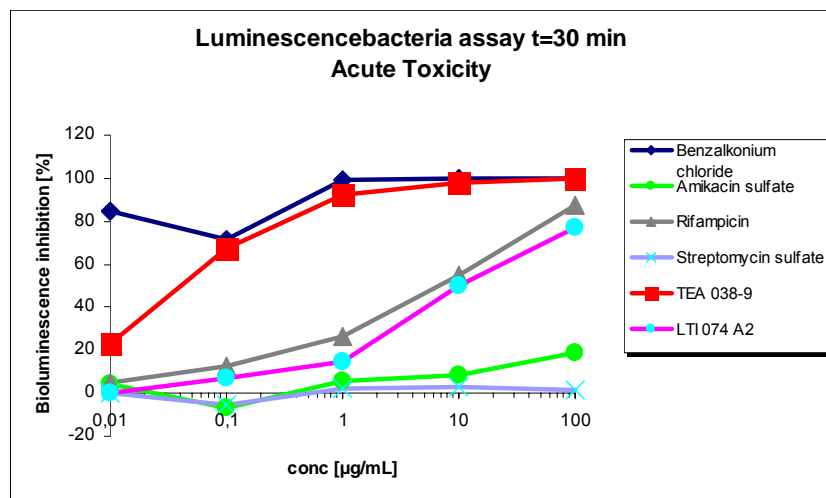


(*2E,9E*)-4-oxooctadeca-2,9-dienoic acid

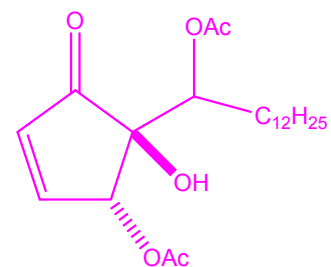


Bioactivity of the isolated fatty acids

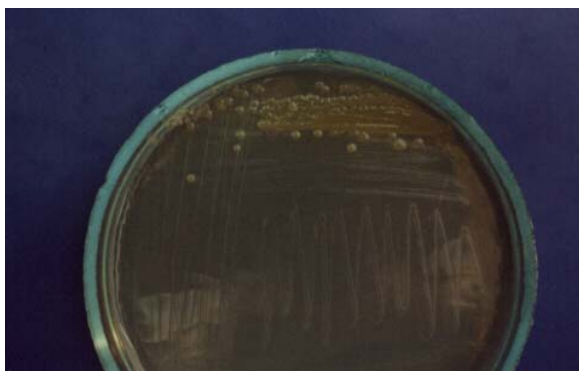
Bactericidal activity: *Vibrio fischeri* (-): 30 min assay



TEA038-9

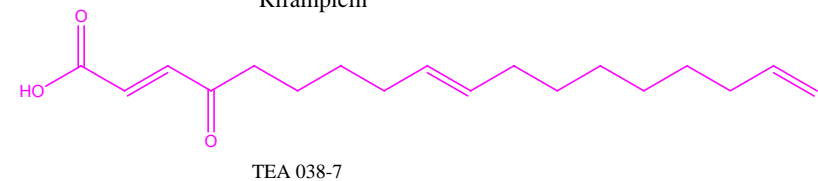
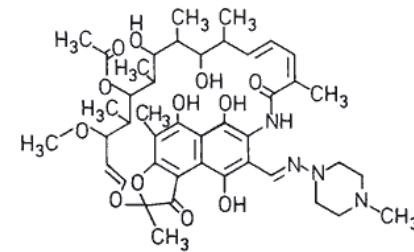
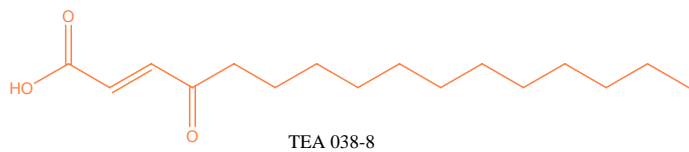
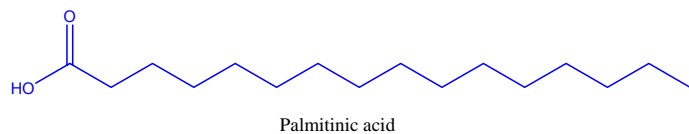
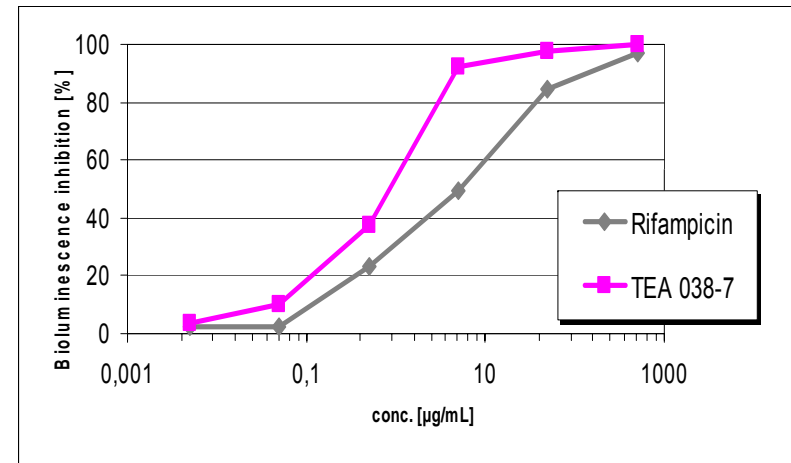
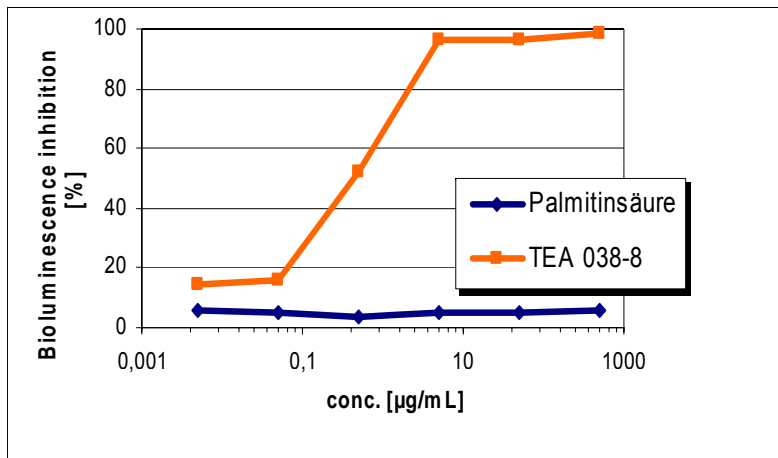


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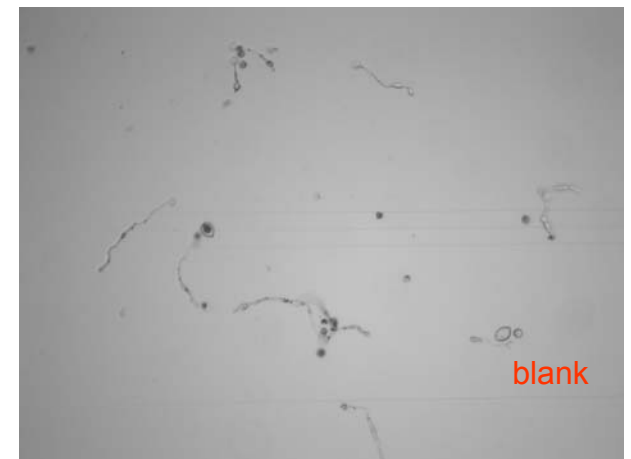
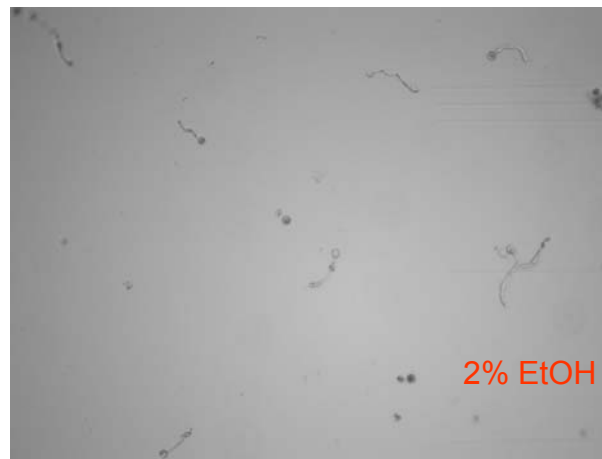
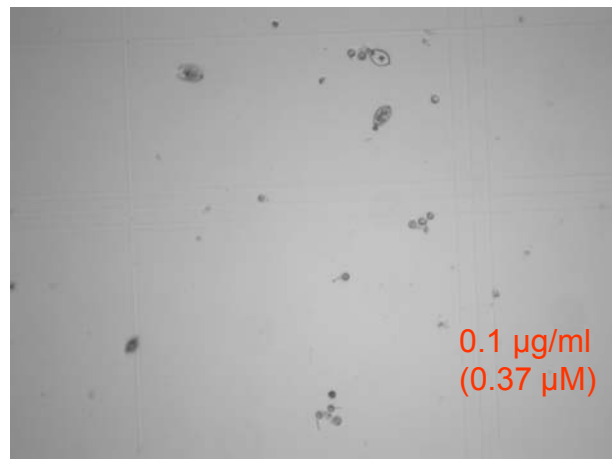
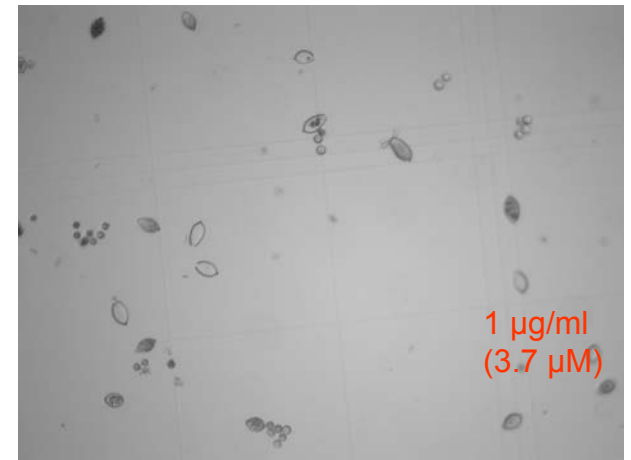
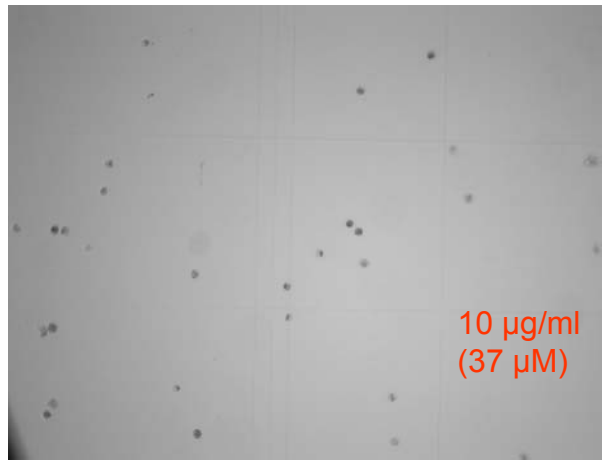
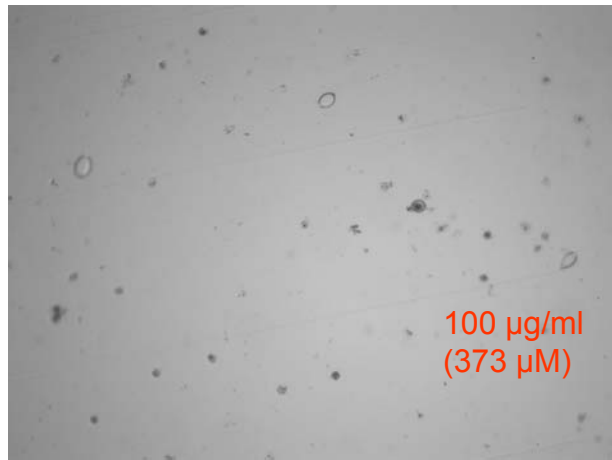
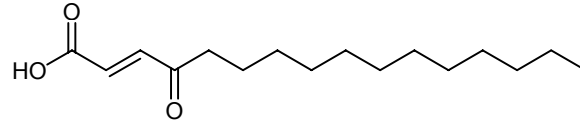
Bioactivity of the isolated fatty acids

Bactericidal activity: *Vibrio fischeri* (-): 24 h assay



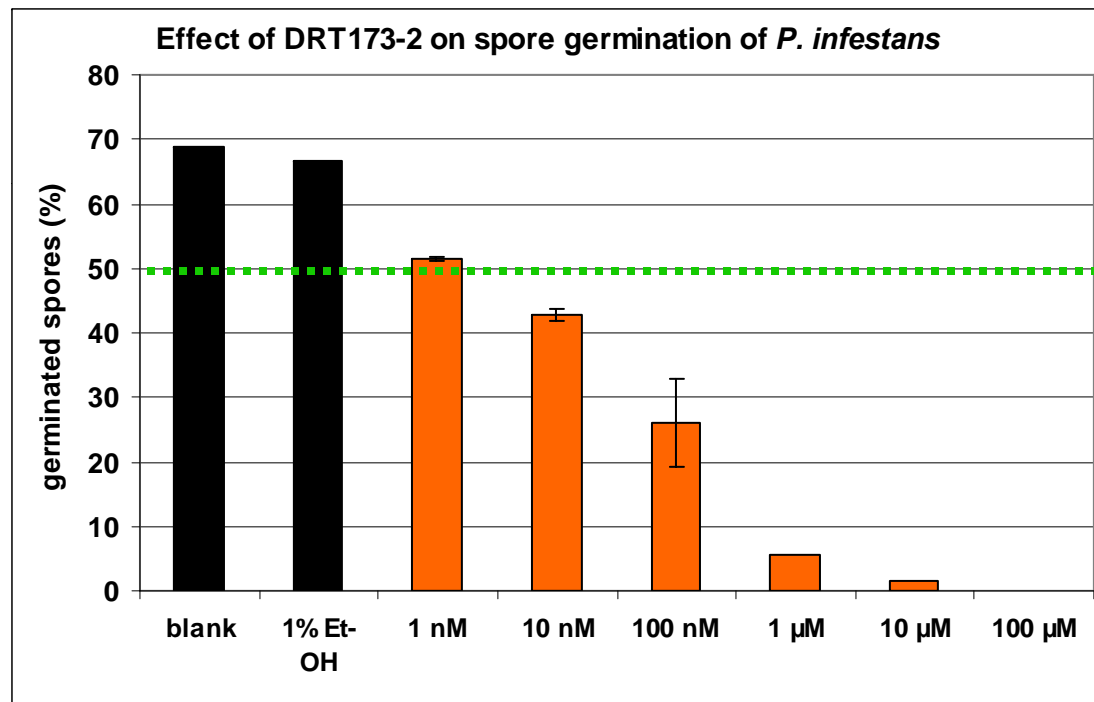
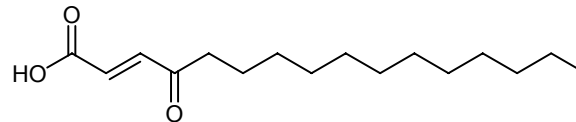
Bioactivity of the isolated fatty acids

P. infestans spore germination assay

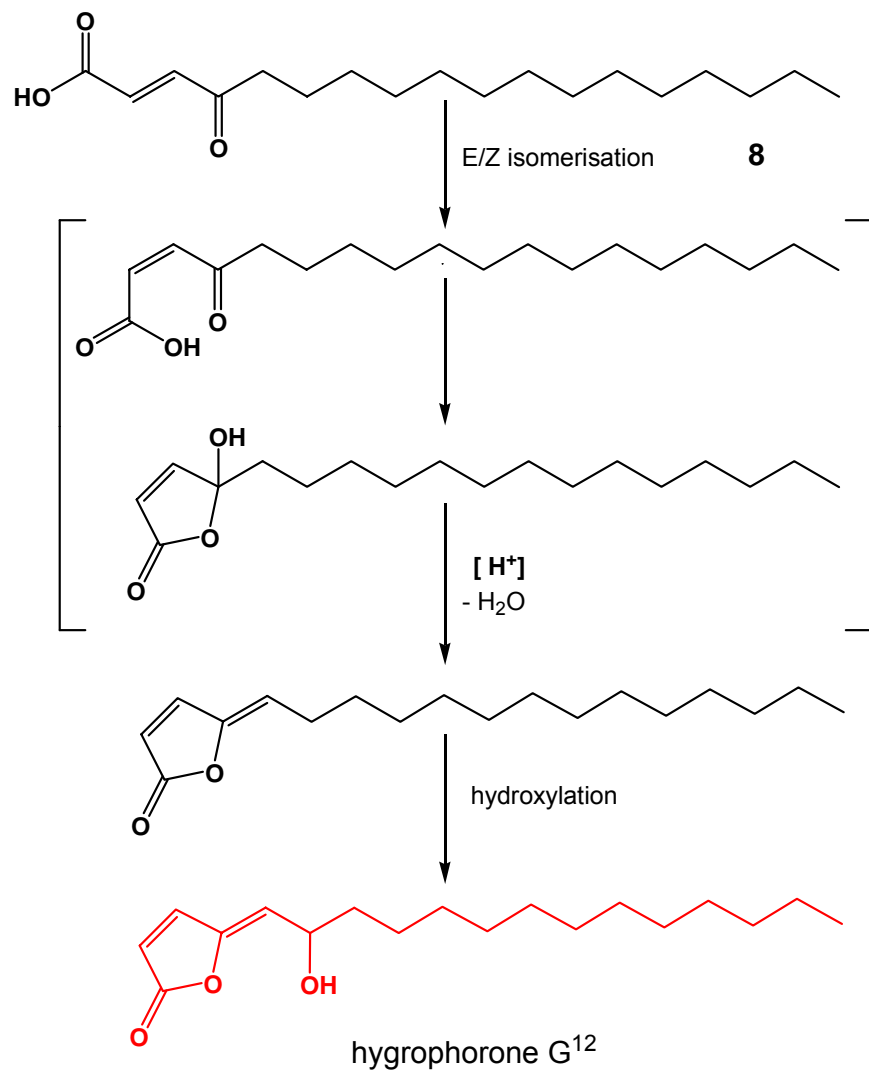


Bioactivity of the isolated fatty acids

P. infestans spore germination assay



Hypothetical relationship of (8) to hygrophorone G¹²



Spotting of the crude extracts or compounds on TLC plates



From fungal fruit body to fungicide - the Strobilurin-story

Strobilurus tenacellus (Pers.) Singer: „pine cone fungus“



From fungal fruit body to fungicide - the Strobilurin-story

Strobilurus tenacellus (Pers.) Singer: „pine cone fungus“



1976: fungicidal properties of *S. tenacellus*
(Prof. T. Anke)

